ClinicSearch

Clinical Oncology Case Reports

Fahim A. Shaltout *

Open Access

Review Article

Good News about the use of the Natural Preservatives in the Meat preservation

Fahim A. Shaltout

Food Control, Faculty of Veterinary Medicine, Benha University, Egypt.

*Correspondence Author: Fahim A. Shaltout, Food Control, Faculty of Veterinary Medicine, Benha University, Egypt.

Received date: September 22, 2025; Accepted date: October 06, 2025; Published date: October 15, 2025

Citation: Fahim A. Shaltout, (2025), Good News about the use of the Natural Preservatives in the Meat preservation, *Clinical Oncology Case Reports*, 4(5): **Doi:**10.31579/2834-5061/027

Copyright: © 2025, Fahim A. Shaltout. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

The Meat and its products are excellent sources of the nutrients for the humans. The Meat and its products also provide a favorable environment for the microbial growth. To prevent the microbiological contamination of the livestock foods, the chemical preservatives, including the nitrites, the nitrates, and the sorbates, have been widely used in the food processing due to their low cost and the strong antibacterial activity. The application of the chemical preservatives is recently being considered by the customers due to the concerns related to the negative health issues. The demand for the natural substances as the food preservatives has increased with the use of the plant origin and the animal origin products, and the microbial metabolites. The natural preservatives inhibit the growth of the spoilage bacteria or the foodborne pathogenic bacteria by increasing the permeability of the microbial cell membranes, the interruption of the protein synthesis, and the cell metabolism. The Natural preservatives can extend the shelf life and inhibit the growth of the bacteria. The natural preservatives can influence the food sensory properties, including the food flavor, the food taste, the food color, the food texture, and the food acceptability. In order to increase the applicability of the natural preservatives, a number of the strategies, including the combinations of different preservatives or the food preservation methods, such as the active packaging systems and the encapsulation, have been used for the applications of the natural preservatives in meat and its products.

Keywords: the meat and its products; the food preservation; the packaging; the foodborne pathogenic bacteria

Introduction

Foodborne pathogenic bacteria, including monocytogenes, the Staphylococcus aureus, the pathogenic Escherichia coli, the Clostridium perfringens, the Campylobacter spp., and the Vibrio spp., cause a large number of the illnesses, with substantial damage to the human health and the economy. The World Health Organization (the WHO), the food contaminated with the foodborne pathogenic bacteria, the chemicals, and the allergens results in 600 million cases of the foodborne illness and four hundred thousand deaths worldwide/ year, Moreover, fifty-six million people die /year and 7.7% of people worldwide suffer from the foodborne diseases. The Meat and its products are essential nutrient sources for the humans due to their excellent protein content, the essential amino acids, the vitamin B groups, and the minerals. The meat and its products also provide an appropriate environment for the spoilage bacteria or the foodborne pathogenic bacteria due to their high-water activity and the nutrient factors (1,2,3,4,5,6 and 7). The food processing has advanced worldwide, resulting in an enhanced the threat of the food contamination by the pathogenic bacteria, the chemical residues, the harmful food additives, and the toxins. The multiplication of the spoilage and the pathogenic bacteria should be controlled to ensure the food safety. The food preservation techniques for protecting the food from the pathogenic bacteria and extending the shelf-life include the chemical methods, such as the use of the preservatives; the physical methods, such as the heat treatment, the drying, the freezing, and the packaging; and the biological methods using the bacteria that have an antagonistic effect on the pathogenic bacteria and produce the bacteriocins. Among them, the addition of the food preservatives that inhibit the growth of the bacteria is a widely used the food protection technique. The countries in the world have different regulations for the food preservatives (8,9,10,11,12,13 and 14). The chemical preservatives have the advantage for the meat production due to the low cost, guaranteed the antibacterial effect or the shelf-life extending activity, and the little effect on the meat taste, the meat flavor, the meat color, and the meat texture. The chemical preservatives tend to be less preferred by the consumers because of a number of health concerns regarding their side effects. The food consumers selected preservatives as the most concerned food additive owing to their negative effects on the health. The Sorbic acid, the benzoic acid, and their salts promote the mutagenic and the carcinogenic compounds. The Nitrites and the nitrate, used as preservative and coloring agents in the meat, have been associated with the leukemia, the colon cancer, the bladder cancer, and others. The Natural preservatives have emerged as alternatives to the chemical preservatives. The Natural preservatives have shown potential to provide the effective antimicrobial activity while reducing the negative health effects. The Meat and its products containing the chemical additives, are a major concern for the public health. The meat processors and the researchers have begun to consider the use of the natural rather than the chemical preservatives (15,16,17,18,19,20 and 21). The 'clean label' food trends, including the meat and its products, began and

Clinical Oncology Case Reports Page 2 of 15

possessed an important source of the food marketing. It includes the consumer, friendly characteristics, such as the chemical additive free, the least processing, a brief list of the food ingredients, and the procedure of the traditional methods. The clean label food material market, including the natural preservatives, is likely to value, mostly owing to growing consumer requests for all the natural products. The natural preservatives such as the nisin, the natamycin, the ε-polylysine, and the grapefruit seed extract are registered, but they are not approved for the meat products, or their concentration is not specified. The replacement of chemical preservatives with the natural preservatives has major positive effects and is being accepted by the customers. The food producers also encounter challenges, including a decrease in price competitiveness due to the relatively high price of the natural preservatives and a decrease in the antibacterial effect due to the food ingredients, such as the carbohydrates, the proteins, and the lipids. In the case of the plant origin substances, the standardization is problematic because of the influence of the country of origin, the soil, and the harvest seasons. The toxicity evaluation or identification of exact compounds for several plant origin compounds contained in extracts and the essential oils have been performed. To solve these problems, various studies have been conducted to optimize the extraction process, combine other antimicrobial substances, apply active packaging, and encapsulate antibacterial substances to improve their utilization (22,23,24,25, 26, 27 and 28).

This review summarizes the current knowledge about the application of the natural preservatives for the meat and its products against the foodborne pathogenic and the spoilage bacteria.

The Application Technique of the Natural Preservatives to the Meat and its Products

The Natural preservatives are manufactured in a variety of formulations including powder formed by drying methods and liquid forms such as essential oils. The Natural preservatives are directly added to the meat products and extend the shelf life by inhibiting the bacterial growth. It is possible to increase the antibacterial effect of the natural preservatives through a combination of the other food processing methods (29,30,31,32,33,34 and 35). In the case of the plant origin natural preservatives, it is necessary to consider the form applied to the food. The Natural preservatives are commonly prepared in the form of extracts using organic solvents, water, and essential oils. The plant extracts obtained from rosemary, chestnut, sage, cranberry, oregano, grape seed, and others have been used as the meat preservatives. The application of the plant origin substances to the meat products in the form of the essential oil because the antibacterial effect of essential oil type is better than that of extract type. It is difficult to apply large amounts of the essential oil to the food because of its distinct organoleptic properties. The Recent developments have attempted to solve this problem by applying essential oils with other antibacterial substances. The advantage of this application is that it reduces the amounts of essential oils with strong flavor and increases antioxidant and antibacterial effects through synergistic effects. In terms of industrial perspective, if the chemical preservatives cannot be completely replaced with the natural preservatives, due to the industrial problems, such as increasing the economic costs or the complexity of the product manufacturing process, they could be replaced gradually by composing a mixed formulation of the chemical preservatives and the natural preservatives (36,37,38,39, 40,41 and 42). The gamma irradiation and the high-pressure processing (the HPP) treatment are the physical foodprocessing methods that can further increase the antibacterial efficacy of the natural preservatives. The Unlike thermal food processing, these two food processing techniques could be used for the pasteurization of the raw meat because it has a minor effect on the food composition. In 1997, the WHO, the Food and Agricultural Organization (FAO), and the International Atomic Energy Agency (the IAEA) concluded that foods processed in the proper doses of the irradiation are nutritionally sufficient and safe to consume. The irradiation is permitted for the food preservation in more than sixty countries. Recent approaches in the food irradiation have involved the use of combined treatments with the natural preservatives to reduce irradiation doses. The gamma irradiation of medium doses (2-6 kGy) with the natural compounds and active packaging has been applied to extend the shelf-life of the meat and its products. The HPP is also a non-thermal technique for the food preservation that inhibits the growth of the bacteria and maintains the natural properties of the food. The HPP is performed under high pressures (100-800 MPa) at the mild temperature or the weak heating. The potential capability of combining the HPP and the natural preservatives including the essential oil and the antibacterial peptides in alleviating both the processing conditions of the HPP and the concentration of the natural preservatives while maintaining antibacterial effects (43,44,45,46,47,48 and 49). The Encapsulation is one of the effective approaches for expanding the applicability of the natural preservatives to the food. The encapsulation was performed with GRAS (generally recognized as safe) materials such as the alginate, the chitosan, the starch, the dextrin, and the proteins using the various techniques including the spray-drying, the extrusion, the freeze-drying, the coacervation, and the emulsification. The application of the natural preservatives to the meat is limited due to their characteristics, such as low solubility and the bioavailability, the rapid release, and the easy degradation. The environmental conditions, such as the pH, the storage temperature and the time, the oxygen and the light exposures could influence the efficacy of the natural preservatives. Through the encapsulation, the natural preservatives, especially hydrophobic compounds (e.g., the essential oil), could improve its stability and expand the versatility of the food processing while maintaining the antibacterial effect (50,51,52,53, 54,55 and 56). The Active packaging is an innovative packaging technology that allows for an interaction with the product and its environment to extend the shelf-life and to ensure its microbial safety while keeping the original properties of the packaged food. In relation to the European Union Guidance to the Commission Regulation, active packaging is a type of the food packaging with a further beneficial function, while providing a protective barrier against the external influence. In the meat processing, the antimicrobial active packaging could be applied in several methods which are the incorporation of the natural preservatives into a sachet inside the packaging, the packaging film composition with the natural preservatives, the packaging coated with the natural preservatives onto the surface of the food, and use the antimicrobial polymers as the packaging materials (57,58,59,60,61,62 and 63). The application of the microorganism origin natural preservatives, known as the bio-preservation, in which the useful bacteria or their antibacterial substances have antagonistic effects on the pathogenic or the spoilage bacteria, are used is also a meat preservation method in the spotlight. This method is mainly involved in the lactic acid bacteria, the Lactobacillus spp., the Leuconostoc spp., the Pediococcus spp., and the Lactococcus spp., that have a GRAS status, widely participate in the fermentation processes, and produce the various antibacterial metabolites such as the organic acids, the hydrogen peroxide, and the bacteriocins. In terms of the application to the meat products, the biopreservation methods included the direct inoculation with the lactic acid bacteria, which has an inhibitory effect on the spoilage or the pathogenic bacteria, the inclusion of the bacterial strains producing the antimicrobial substances in the fermentation starter, and the treatment with the purified bacteriocins (64,65,66,67,68,69 and 70).

The Natural Preservatives from the Plants and Their Application for the Meat and its Products

The antibacterial effect of the plant origin natural preservatives is closely related to the polyphenols, the phenolics, and the flavonoids. The Plant origin polyphenols have various classifications and structures, as the phenolic acids (the caffeic acid, the rosmarinic acid, the gallic acid, the ellagic acid, the cinnamic acid), the flavones (the luteolin, the apigenin, the chrysoeriol), the flavanols (the catechin, the epicatechin, the epigallocatechin, the gallocatechin, and their gallate derivatives), the flavanones (the hesperidin, the hesperetin, the heridictyol, the naringenin), the flavonols (the quercetin, the kaempferol, the myricetin), the isoflavones (the geinstein, the daidzin, the formononetin), the coumarins (the coumarin, the warfarin, the 7-hydroxycourmarin), the anthocyanins (the pelagonidin, the delphinidin, the cyanidin, the malvidin), the quinones (the naphthoquinones, the hypericin), the alkaloids (the caffeine, the berberine, the harmane), and the terpenoids (the menthol, the thymol, the lycopene, the capsaicin, the linalool) (71,72,73,74,75,76 and 77). The Polyphenols have been recognized for their effective antimicrobial

Clinical Oncology Case Reports Page 3 of 15

properties. Although the antimicrobial mechanism has not yet been clearly elucidated, the cell membrane-disturbing molecules, such as the hydroxy group (OH-), which induces the leakage of intracellular components, inactivation of metabolic enzymes, and extinction of the adenosine triphosphate (the ATP) structure; direct pH change in the environment by the improvement in proton concentration, reduction of the intracellular pH by separation of acid molecules, and modification of the bacterial membrane permeability; an organic acid in the plant extracts may influence the oxidation of the nicotinamide adenine dinucleotide (the NADH), the eliminating, the reducing agent used in the electron transport system (78,79,80,81,82,83 and 84).

The Rosemary

The Rosemary (the Rosmarinus officinalis L.) is a perennial herb with the woody, the aromatic, and the evergreen needle-like leaves. Originally from the Mediterranean region, it is broadly distributed throughout the globe. The Rosemary has been used as a spice and the flavoring agent in the food. The Rosemary essential oil is known to contain fifteen kinds of the bioactive compounds. The principal compound was 1,8-cineole (35.32%). Other major compounds were the camphor, the α -pinene, the transcaryophyllene, the α -thujone, and the borneol (85,86,87,88,89,90 and 91). The antibacterial effect of the rosemary ethanol extracts against the Listeria monocytogenes in the beef. The application of 45% rosemary ethanol extract for the Listeria monocytogenes on the beef led to a two-log colony forming unit / gram reduction in the incubation at 4 °C for nine days. In the chicken meat, the effect of the rosemary essential oil on the inhibition of the Salmonella Enteritidis and the spoilage protective effects at 4 and 18 °C was investigated. The 5 mg/mL of the rosemary essential oil induced the decrease in the coliform, the aerobic bacteria, the lactic acid bacteria, and the anaerobic bacteria at 18 °C for one day. The Comparing with the untreated chicken meat, the reductions of 1.75 log Colony Forming Unit / gram (the coliform), 0.87 log Colony Forming Unit / gram (the aerobic bacteria), 1.05 log Colony Forming Unit / gram (the lactic acid bacteria) and 1.28 log Colony Forming Unit / gram (the anaerobic bacteria) were observed in the group treated with rosemary essential oil at 18 °C. The Rosemary oil reduced the S. Enteritidis by more than two log Colony Forming Unit / gram at 18 °C, but less than one log Colony Forming Unit / gram at 4 °C (92,93,94,95,96,97 and 98). The rosemary essential oil applied with modified atmosphere packaging for the inhibition of the foodborne pathogenic bacteria (the S. Typhimurium and the Listeria monocytogenes) in the poultry filets under the refrigerated conditions for seven days was examined. The 0.2% rosemary essential oil did not affect the sensory profile and inhibited the growth of both the pathogenic bacteria in the laboratory media within one day. The Treatment with 0.2% rosemary essential oil did not affect the reduction in the S. Typhimurium, but showed weak antibacterial activity against the Listeria monocytogenes until the first day of the storage (0.1 log Colony Forming Unit / gram the reduction compared to the control) (99,100,101,102,103,104 and 105).

The Sage

The Sage (the Salvia officinalis L.), belonging to the Lamiaceae family, has been used since prehistoric eras because of its flavor, taste, therapeutic, and preservative properties. The Sage is known to contain considerable amounts of the rosemary acid, the p-coumaric acid, and the benzoic acid. The Sage essential oils, the camphor, the carvacrol, the R (+) limonene, and the linalool are the major components in terms of content (106,107,108, 109,110 and 111). The antibacterial effects of various sage preparations were assessed for low-pressure mechanically separated meat in vacuum packaging stored at -18 °C for nine months. The mechanically separated meat from the chickens with the addition of the sage extracts inhibited the growth of all groups of the bacteria (the mesophilic aerobic bacteria, the psychrotrophic bacteria, the Enterobacteriaceae, the coliforms, and the enterococci). The most effective antibacterial effect was exhibited by the 0.1% sage essential oil-treated (112,113,114,115,116 and 117). The antibacterial effect of the sage essential oil (0.625%) on the survival of the Listeria monocytogenes in the Sous-vide cook-chill beef stored in the refrigerated storage (2 or 8 °C) for 28 days. The decrease of one log Colony Forming Unit / gram of the Listeria monocytogenes was detected in the sage essential oil-treated groups compared to the control at 2 $^{\circ}$ C. Although the exponential growth was observed from the day 14, decrease the Listeria monocytogenes counts of one log Colony Forming Unit / gram were detected in the sage essential oil treated samples stored at 8 $^{\circ}$ C (118,119,120,121,122 and 123).

The Thyme

The Thyme (the Thymus vulgaris) is a representative herb used together with the meat and its products. The application of the thyme in the meat products can elevate the antioxidant, the antibacterial, the shelf-life extension, and the sensory properties. In the meat sausage, the thyme essential oil inhibited 2.69 log Colony Forming Unit / gram of coagulasepositive Staphylococcus and 4.41 log Colony Forming Unit / gram of aerobic mesophilic bacteria, respectively, at a concentration of 0.95% by mixing with 1% (w/w) powdered beet juice. The sensory properties, odor, flavor, and overall acceptability improved (124,125,126,127,128 and 129). The 1% thyme oil led to the reduction in the S. enterica by three log Colony Forming Unit / gram during the margination process with lemon juice and 0.5% Yucca schidigera extract in the raw chicken breast. The major composition of the thyme oil revealed 51.1% and 24.1% thymol and Ocymene, respectively. The antibacterial effects of thyme may be due to additive or synergistic effects with its major and/or minor components. The Thymol and its synergistic effect with other phenolic compounds, such as the carvacrol, the p-cymene, and the γ-terpinene, can change the permeability of the bacterial cell wall, leading to cell death (130,131,132,133,134 and 135). The Thyme essential oil encapsulated with the casein and the maltodextrin was evaluated for its antibacterial potential in the vitro and in the situ (the hamburger-like meat products). The encapsulated thyme essential oil showed the same minimum inhibitory concentration (0.1 mg/mL) against the Escherichia coli, the S. Typhimurium, the Staphylococcus aureus, and the Listeria monocytogenes as that of the unencapsulated thyme essential. In the treated groups with 1% (v/v) of the encapsulated thyme essential oil for the meat, the Escherichia coli counts were decreased from 23 most probable number (the MPN)/ gram to 0 MPN/ gram, which was similar to the conventional preservative (the sodium nitrate) used as a control until 14 days of the refrigerated storage (4 °C) (136,137,138,139,140 and 141).

The Oregano

The Oregano (the Origanum vulgare) is regularly used in the Mediterranean foods. The oregano essential oil has recognized antibacterial and antioxidant properties for the extension of the shelf-life. The antibacterial effects of the oregano were due to two bioactive the polyphenols, the thymol and the carvacrol (142,143,144,145,146 and 147). The oregano essential oil and its effect on the shelf life of the black wildebeest Biceps femoris muscles was investigated at 2.6 °C. The components of the oregano oil were the thymol, the carvacrol, the pcymene, the β -caryophyllene, the γ -terpinene, the α -humulene, and the α pinene; among them, the carvacrol (42.94%) and the thymol (17.40%) were the highest. The total viable counts and the lactic acid bacteria reached the spoilage limit (seven log Colony Forming Unit / gram) after three days. The growth rates for the total viable counts and the lactic acid bacteria in the treated group were 40% higher than those in the untreated groups (148,149,150,151,152 and 153). The combinatorial effect of the oregano essential oil with the caprylic acid was studied in the vacuumpacked minced beef. The addition of 0.2% oregano essential oil with 0.5% caprylic acid and 0.1% citric acid in the minced beef reduced the counts of the lactic acid bacteria by 1.5 log Colony Forming Unit / gram in vacuum packaging. The cell counts of the psychrotrophic bacteria and the Listeria monocytogenes were reduced by more than 2.5 log Colony Forming Unit / gram at 3 °C for 10 days. The Oregano essential oil inhibits the growth of the bacteria by releasing volatile components during the drying process. The addition of the oregano essential oil composed of the carvacrol (64.5%), the p-cymene (5.2%), and the thymol (2.9%) inhibited the S. Enteritidis and the Escherichia coli in the beef drying process. For drying, a filter paper was soaked with oregano essential oil and placed in front of the fan of the drier. The beef samples were dried at 55 °C for 6 hours. Both the bacteria (the S. Enteritidis and the Escherichia coli) were not detected after treatment with three mL of oregano essential oil (154,155,156,157,158 and 159).

Clinical Oncology Case Reports Page 4 of 15

The Chestnut

The Castanea crenata was classified into the Castanea family and is a woody plant native to the East Asia, including the Korea and the Japan. The Castanea sativa is one of the most important Castanea families and the food resources of the European areas for long periods. Chestnut shells contain abundant phenols and hydrolyzable tannins. The chestnut inner shell extracts using ethanol exhibited antimicrobial effects against C. jejuni in the chicken meat at a concentration of two mg/mL. The polyphenol and flavonoid contents of chestnut inner shell ethanol extracts were 532.96 \pm 3.75 mg gallic acid/100 g and 12.28 \pm 0.03 mg quercetin/100 g, respectively (160,161,162,163,164 and 165). The influence of the chestnut extracts (the Castanea sativa) on the leaf, the bur, and the hull of the beef patties under refrigerated conditions (2 ± 1 °C) for 18 days to extend the shelf-life. Inside the chestnut extracts from the leaf, the bur, and the hull, only the leaf extract at a concentration of 1000 mg/kg had weak antimicrobial activity. The lactic acid bacteria and the Pseudomonas spp. were reduced by 0.37 log Colony Forming Unit / gram and 0.33 log Colony Forming Unit / gram at seven days, respectively (166,167,168,169,170 and

The Grapefruit Seed Extract

The Grapefruit Seed Extract is a byproduct of the Citrus paradise. The Grapefruit Seed Extract contains the various phenolic compounds and the flavonoids, such as the catechin, the citric acid, the naringenin, the procyanidin, and the epicatechin gallate. The Grapefruit Seed Extract has been described to have a wide-ranging spectrum antimicrobial, the antiparasitic, and the antifungal activities. The Polyphenols in the Grapefruit Seed Extract are unstable but can be chemically modified to become more stable using quaternary ammonium compounds, such as the benzethonium chloride, during the industrial procedure of the commercial Grapefruit Seed Extract preparations (172,173,174,175,176 and 177). The bacteriostatic effect of the commercial Grapefruit Seed Extract (the Citricidal) on the sous-vide chicken products against the Clostridium perfringens. The cell numbers of the Clostridium perfringens were consistently 2.5 log Colony Forming Unit / gram regardless of the treatment or the control groups until 9.5 h of stored at 19 °C; the storage of the control and 50 or 100 ppm Grapefruit Seed Extract treated groups at 25 °C for more than six hours resulted in fast growth rates of the Clostridium perfringens, showing 2–3 log Colony Forming Unit / gram. The Grapefruit Seed Extract concentrations at 200 ppm inhibited the growth of the Clostridium perfringens stored at 19 and 25 °C. The active packaging system for the inhibition of the foodborne pathogenic bacteria used the mixed natural preservatives consisting of the Grapefruit Seed Extract (80 mg/m2) with the cinnamaldehyde (200 mg/m2) and the nisin (60 mg/m2) was assessed for the beef storage. The active packaging showed decrease the counts of the psychrotrophic and the anaerobic bacteria compared to the control groups at 1-2 log Colony Forming Unit / gram. The packaged beef samples with mixed natural preservatives showed a decrease in the Listeria monocytogenes, the Staphylococcus aureus, and the C. jejuni for 4.7 log Colony Forming Unit / gram, 0.81 log Colony Forming Unit / gram, and 3.1 log Colony Forming Unit / gram compared to wrapped packaging at 28 days of the refrigerated storage, respectively. The C. jejuni was observed below the detection limit after 21 days of the storage (184,185,186,187,188 and 189)

The Cinnamon

The Cinnamon is a native plant in Asia that is acquired from the inner bark of the genus Cinnamonum. The Cinnamon contains several active compounds, such as the cinnamaldehyde, the eugenol, the cinnamyl acetate, the L-borneol, the β -caryophyllene, the caryophyllene oxide, the camphor, the L-bornyl acetate, the α -terpineol, the α -cubebene, the α -thujene, and the terpinolene. The cinnamon (Cinnamonum cassia) essential oils could inhibit the L. monocytogenes in the ground beef at the refrigerated (0 and 8 °C) and the frozen (–18 °C) conditions. The concentration of five percentage cinnamon essential oil to decrease by 3.5–4.0 log Colony Forming Unit / gram of Listeria monocytogenes at 0 and 8 °C for seven days. Under the frozen conditions, the Listeria monocytogenes was reduced by 3.5–4.0 log Colony Forming Unit / gram over 60 days. The antibacterial effect and the shelf-life extending activity

were evaluated using a chitosan edible coating containing 0.6% cinnamon essential oil on the roast duck slices under the modified atmosphere packaging (30% carbon dioxide (CO2)/70% nitrogen (N2)) at the storage at 2 ± 2 °C for 21 days. The edible coating with cinnamon essential oil showed the total viable counts reduced by one log Colony Forming Unit / gram compared to the control after 14 days of the storage. It is similar to the results of the Enterobacteriaceae counts. The number of the lactic acid bacteria was decreased than that of the control until the day 7 of the storage, but there was no significant difference from day 11 of the storage. The growth of the Vibrio spp. was delayed using the edible coating with the cinnamon essential oil within the earlier period of the storage as a result of the microbial diversity sequencing (196,197,198, 199, 200 and 201).

The Turmeric

The Turmeric (Curcuma longa L.) has long been used as a flavor and the color agent in the food and the traditional medicine to treat the various diseases, mainly in the South and East Asia. The main active compounds of the turmeric originate from its constituents, called the curcuminoids. The Curcuminoids (the curcumin, the demethoxycurcumin, and the bisdemethoxycurcumin) content of the turmeric varies between about 2–9% based on its growth environments, such as the cultivar, the soil, and the climatic conditions. The antibacterial effect of the turmeric on the chicken breast meat was assessed for the Escherichia coli and the Staphylococcus aureus stored at 4 °C for two days. When 1% turmeric powder was added, no difference in the Staphylococcus aureus counts was observed between the turmeric treated and the control groups. In the case of the Escherichia coli, a reduction of 0.2 log Colony Forming Unit / gram was observed, but this was not statistically significant (202,203,204,205, 206 and 207). The chicken meat was treated with the turmeric powder and the gamma irradiation to improve the meat quality and stability. The total aerobic bacteria and the coliforms were completely decontaminated with 3% turmeric powder and 2 kGy of the gamma irradiation at 4 °C for 14 days. The microbial characteristics of the edible coatings using the turmeric starch and the bovine gelatin were examined in the frankfurter sausages. The edible coating was developed with a 5% (w/w) aqueous solution of the turmeric starch and the gelatin. The microbial growth of the coated sausages stored at 5 °C for 20 days decreased by 2.21, 1.01, and 1.65 log Colony Forming Unit / gram for the mesophilic bacteria, the lactic acid bacteria, and the psychotropic bacteria, respectively. At 10 °C, the decreases were 1.57, 2.14, and 1.99 log Colony Forming Unit / gram for the mesophilic bacteria, the lactic acid bacteria, and the psychotropic bacteria, respectively (208,209,210,211,212 and 213).

The Plant origin Antimicrobial Peptides

The Plant origin the Antimicrobial Peptides have been studied for their potential to inhibit the different pathogenic bacteria, including the food spoilage bacteria, the food poisoning bacteria, the mold, and the yeast species. The antibacterial peptide Leg1 from the chickpea legumin were reported in the meat application of the plant origin the Antimicrobial Peptides. The Raw pork was pretreated with Leg1 and inoculated with the Escherichia coli and the B. subtilis. The bactericidal activity was measured at 37 °C for 16 hours. The minimum bactericidal concentrations of Leg1 on the pork were 125 μ M and 15.6 μ M for the Escherichia coli and the B. subtilis, respectively. This was the same concentration as the MBC of the nisin, the bacteriocin from the Lactococcus lactis, for the tested strains. The Antimicrobial Peptides from pea (the 11SGP) and the red kidney bean (the RBAH) were used to extend the shelf life of the raw buffalo meat. In the laboratory media, the Gram positive (the L. monocytogenes, the B. cereus, and the Streptococcus pyogenes) and the Gram-negative (the Escherichia coli, the Pseudomonas aeruginosa, the Acinetobacter baumannii) bacteria were inhibited by 11GSP (60 µg/mL) and the Gramnegative bacteria by 60% and the Gram-positive bacteria by 90%. RBAH (60 μg/mL) alleviated the growth of the Gram-negative bacteria by 56% and the Gram-positive bacteria by 85%. In the buffalo meat, the counts of the mesophilic bacteria of 11SGP (400 µg/gram) and the RBAG (400 µg/ gram) treated groups decreased by 1.60 log Colony Forming Unit / gram and 1.94 log Colony Forming Unit / gram compared to the control groups. The psychrophilic bacteria, 11SGP and the RBAG reduced by 1.10 log Colony Forming Unit / gram and 1.47 log Colony Forming Unit / gram,

Clinical Oncology Case Reports Page 5 of 15

respectively, after 15 d of the refrigerated storage (4 $^{\circ}$ C) (172,173,174,175 and 176).

The Natural Preservatives from the Animals and Their Application for the Meat and its Products

The Various antibacterial systems of the animal sources are associated with the defense mechanisms against external intruders. The preservatives derived from the animal sources include the lysozymes, the lactoferrin, the ovotransferrin, the lactoperoxidase, the Antimicrobial Peptides from the livestock animals, and the polysaccharides. The Lysozyme can suppress several Gram-positive bacteria because of the Lysozyme distinctive ability to injure bacterial membranes by hydrolyzing the 1,4-β-linkage between the N acetyl D glucosamine and the N acetyl muramic acid of the peptidoglycan in the bacterial membrane. The Peptide based antibacterial substances containing the Antimicrobial Peptides from the animal sources, the ovotransferrin, and the lactoferrin could influence the cell membranes or the synthesize Antimicrobial Peptides, the peptides, and the enzymes. The antibacterial mechanism of the Antimicrobial Peptides due to the attachment to the bacterial cell membrane and disturb its integrity, resulting in the cell lysis. The Antimicrobial Peptides may also exert more complex activities that inhibit the metabolic and the translational systems. The ovotransferrin isolated from the eggs increased the cell membrane permeability of the Gram positive and the Gram-negative bacteria. The ovotransferrin destroyed the cell membrane integrity, increased the permeability of the pathogen membranes, and induced morphological changes. The Lactoferrin has antibacterial effects related to the large cationic patches present on the surface and the iron impoverishment. The Lactoferrin has an antibacterial effect only when in its iron free state and the iron saturated lactoferrin has a limited antimicrobial activity. The Lacroperoxidase oxidizes the sulfhydryl groups of the proteins present in the bacterial membrane, which could be injured by the efflux of the potassium ions, the amino acids, the peptides, and the enzymes (177,178,179,180,181 and 182).

The Lysozyme

The Lysozyme (the muramidase or the N acetyl muramichydrolase) is mainly extracted from the hen egg whites and is known as an antimicrobial enzyme. The Lysozyme is a glycoside hydrolase that hydrolyses the linkages in the peptidoglycan at the Gram-positive bacterial cell wall. The Lysozyme is composed of 129 amino acids, which contain the disulfide bonds and the tryptophan, the tyrosine, and the phenylalanine residues. The Lysozyme has been used commercially, named the Inovapure, against the spoilage and the foodborne pathogenic bacteria to prolong the shelf life of the raw and the processed meat. The Modified lysozyme, the high hydrophobicity, and the low hydrolytic activity compared to the lysozyme monomer, at the concentrations of 5%, exhibited low microbial growth rates (the total viable count 4.59 log Colony Forming Unit /cm2; the molds and the yeasts 2.17 log Colony Forming Unit /cm2) in the pork surface with the modified atmosphere packaging with composites of 50% O2, 40% CO2, and 10% N2. The mixed antimicrobials consisting of the lysozyme ppm), the nisin (250 ppm), and the ethylenediaminetetraacetic acid (the EDTA) (20 mM) had antibacterial effects against the Listeria monocytogenes, the total viable counts, the Enterobacteriaceae, the Pseudomonas spp., and the lactic acid bacteria in the ostrich meat patties with the air and the vacuum packaging. The mixed lysozyme preparations reduced the Listeria monocytogenes below the official detection limit of the European Union (<2 log Colony Forming Unit / gram) in the ostrich meat patties. The treated samples showed a decrease in the total viable counts by one log Colony Forming Unit / gram after two days of the storage and tended to increase thereafter. The Enterobacteriaceae and the Pseudomonas spp. we're not affected by the mixed antimicrobials in either the packaging atmosphere, and the reduction in the lactic acid bacteria was detected at two log Colony Forming Unit / gram. The combination of the lysozyme with the chitooligosaccharide presented a more effective antibacterial effect against the Gram-negative bacteria than the lysozyme alone. In the minced mutton, the mixture of the lysozyme and the chitooligosaccharide led to complete removal of 3-4 log Colony Forming Unit / gram of the inoculated Escherichia coli, Pseudomonas fluorescens, and B. cereus during four hours at the ambient temperature. The Staphylococcus aureus was not completely eliminated, but was reduced up to two log Colony Forming Unit /gram (183,184,185,186 and 187)

The Ovotransferrin

The Egg white contains 13% ovotransferrin (the conalbumin), which is a monomeric 77.9 kDa glycoprotein comprised of 686 amino acid residues. The ovotransferrin contains N and C globular parts, each of which can reversibly Fe3+ and CO32-. The ovotransferrin is the main constituent of the egg's defense system for the bacteria, as it renders the iron unusable for the microbial growth within the albumen. The antimicrobial effects of the ovotransferrin against the Escherichia coli in the fresh chicken breast involved in κ carrageenan film. The growth of the Escherichia coli in the fresh chicken breast wrapped with the active film was 2.7 log Colony Forming Unit / gram by the addition of 25 mg of the ovotransferrin in combination with 5 mM EDTA. The ham models, 25 mg/mL of ovotransferrin with 100 mM sodium bicarbonate (NaHCO3) did not show any antibacterial effects against the Escherichia coli O157:H7 and the Listeria monocytogenes in commercial hams, whereas 25 mg/mL ovotransferrin with half percentage citric acid had bacteriostatic effects against Listeria monocytogenes (188,189,190,191,192 and 193).

The Lactoferrin

The Lactoferrin, a glycoprotein that belongs to the transferrin protein family in the milk and the milk products as well as the neutrophil granules and exocrine secretions in the mammals, was able to bind the iron within the cells. The ability of this 80 kDa protein to control free iron levels contributes to its bacteriostatic and the health beneficial characteristics, such as stimulating bone growth, protecting the intestinal epithelium, and promoting the immune system in animals. In the ground beef, application of the active lactoferrin, the immobilized lactoferrin with the glycosaminoglycans, and solubilized in the citrate/bicarbonate buffer systems at concentrations of three percentage and five percentage resulted in two log Colony Forming Unit / gram reductions of Escherichia coli O157:H7 at 10 °C for nine days. The reduction of the S. Enteritidis growth was 0.8 log Colony Forming Unit / gram when the active lactoferrin concentration was increased to two-point five percentage. A single application of half percentage active lactoferrin reduced Listeria monocytogenes in the beef, resulting in two log Colony Forming Unit / gram. The Bovine lactoferrin (half mg) was tested against the Escherichia coli O157:H7 and P. fluorescens inoculated on the chicken with HPP treatments between 200 and 500 MPa for 10 min at 10 °C. As a result, the P. fluorescens was decreased when the lactoferrin was combined with the HPP treatment at 300 MPa for 2.3 log Colony Forming Unit / gram additional reduction compared to only 300 MPa treatment on day 9. Additional reductions in the Escherichia coli O157:H7 counts obtained by combined treatments remained below 0.5 log Colony Forming Unit / gram. (194,195,196,197 and 198)

The Lactoperoxidase

The Lactoperoxidase is a member of the peroxidase family. It is a ubiquitous active enzyme in bovine milk, which has antimicrobial effects. Bovine lactoperoxidase is a glycoprotein that contains a peptide chain of 78.4 kDa and catalyzes the oxidation of thiocyanate ions (SCN) in lactoperoxidase, producing oxidizing products, such as hypothiocyanite and hypothiocyanous acid. The lactoperoxidase coated with alginate at concentrations of 2, 4, and 6% on the shelf life of the chicken breast filets. The chicken samples with active coating of alginate and 6% lactoperoxidase showed a reduction of the Enterobacteriaceae, the P. aeruginosa, and the aerobic mesophilic bacteria by five log Colony Forming Unit / gram, 4 log Colony Forming Unit / gram, and 2.5 log Colony Forming Unit / gram at 16 days of the refrigerated storage, respectively. The antimicrobial effects of the lactoperoxidase were also assessed against the Listeria monocytogenes and the S. Enteritidis in the sliced dry cured ham for 60 d at 8 °C treated with the HPP at 450 MPa. The synergistic effect of lactoperoxidase and pressure was confirmed as the S. Enteritidis decreased below the detection limit (one log Colony Forming Unit / gram). The Listeria monocytogenes, the synergistic effect reduced cell viability by 0.86 log Colony Forming Unit / gram compared

Clinical Oncology Case Reports Page 6 of 15

with the untreated samples at the end of the storage. In the beef, the effect of the lactoperoxidase on the growth of the inoculated pathogenic bacteria (four log Colony Forming Unit / gram) composed of the Staphylococcus aureus, the Listeria monocytogenes, the Escherichia coli O157:H7, the S. Typhimurium, the P. aeruginosa, the Yersinia enterocolitica, and the indigenous microbiota was investigated. The pathogenic bacteria used in the experiment were reduced compared to the control at a chilling regime (–1 to 12 °C) for 42 days. The total aerobe and the Pseudomonas spp. increased less in the lactoperoxidase treated group than in the control group, but the antibacterial effect was not exhibited for anaerobes and the lactic acid bacteria (199,200,201,202 and 203).

The Livestock Animal origin

The Livestock animal origin products have been used as a source of the Livestock animal origin products. Among these byproducts of the livestock, the blood, the bones, the collagen, the gelatin, the liver, the lungs, the placenta, the skin, and the visceral mass are important sources of the Livestock animal origin products, as well as the muscle parts. The bovine cruor, a slaughterhouse byproduct containing mainly hemoglobin, broadly described as a rich source of fibrin proteins, was investigated for the extraction of the Livestock animal origin products. The faction named α137-141 (polypeptide with five components, Thr Ser Lys Tyr Arg), a small (0.65 kDa), and hydrophilic Livestock animal origin products deviated from hemoglobin. The a137-141 preservative (0.5%, w/w) had bacteriostatic effects on the total microbial population, coliform bacteria, the yeasts, and the molds at 4 °C for 14 d on the beef. The Livestock animal origin products isolated from porcine leukocytes had antibacterial effects on the proliferation of Staphylococcus aureus and Escherichia coli inoculated in the ground meat (the boneless ham) and the sausage minces. The 20 µg/ gram Livestock animal origin products decreased by 1.3 log Colony Forming Unit / gram of Staphylococcus aureus and 1.5 log Colony Forming Unit / gram of the Escherichia coli in ground meat. It was also achieved that 160 µg/ gram of the Livestock animal origin products had the best inhibition and decreased in 3.9 log Colony Forming Unit / gram of the Staphylococcus aureus and 3.3 log Colony Forming Unit / gram of the Escherichia coli at 6 hours in the ground meats. In sausage mince, the Livestock animal origin products at concentrations of 160 µg/ gram could decrease by three log Colony Forming Unit /g of Staphylococcus aureus and 2.7 log Colony Forming Unit / gram of the Escherichia coli at 12 hours. After one day of the storage, no visible colonies of the Staphylococcus aureus or the Escherichia coli were detected in the sausage mince (203,204,205,206,207 and 208).

The Natural Preservatives from the Microorganism and Their Application for the Meat and its Products

The Lactic acid bacteria strains secrete several bacterial growth inhibitory substances (the organic acids, the diacetyl, the phenyl lactate, the hydroxyphenyl lactate, the cyclic dipeptides, the hydroxy fatty acid, the propionate, and the hydrogen peroxide), the bacteriocins (the nisin, the acidophilin, the bulgaricin, the helveticin, the lactacin, the pediocin, the plantarim, the diplococcin, and the bifidocin), and the bacteriocin like inhibitory substances (the Bacteriocin Like Inhibitory Substance), which exhibit antibacterial activity and can control the spoilage and the foodborne pathogenic bacteria. The various bacteriocins, commercial bacteriocin preparations have been applied using nisin and pediocin. The Bacteriocins are peptides or proteins with antibacterial and antifungal effects that produce bacteria, mainly the lactic acid bacteria. The compounds are considered potential natural preservatives because of their inhibitory effects on the food spoilage or pathogenic bacteria. The Lactic acid bacteria bacteriocins vary in accordance with molecular size, the chemical structure, modifications during biosynthesis, presence of modified amino acid residues, and antimicrobial mechanisms. The Lactic acid bacteria bacteriocins can be categorized into two major classes: class I (the lanthionine containing antibiotics) with three subclasses (the Ia, Ib, and Ic) and the class II with four subclasses (IIa, IIb, IIc, and IId). The Class I bacteriocins generally include 19–50 amino acid residues (<5 kDa) and are largely post translationally modified, ensuring the nonstandard amino acids, such as the lanthionine, the β methyllanthionine, and the labyrinthine. The class I bacteriocins are further subdivided into the class

Ia (the lantibiotics), the class Ib (the labyrinthopeptins), and the class Ic (the sanctibiotics). The Class II bacteriocins comprise the small, heat stable, the non-modified peptides (<10 kDa). It can be further subdivided into the class IIa (the pediocin like bacteriocins), the class IIb (the nonmodified bacteriocins with two or more peptides), the class IIc (the circular bacteriocins), and the class IId (the non-pediocin like bacteriocins). The Pediocin like bacteriocins (the class IIa) can be regarded as the main subgroup among all classified the Lactic acid bacteria bacteriocins. The Class III bacteriocins are classified as the high molecular weight (>30 kDa) and the thermally unstable peptides. The Class IV bacteriocins are large peptides complexed with the lipids or the carbohydrates. The bacterial cell surface exhibits a negative charge because the anionic characteristics of the cell membrane consist of the phosphatidylethanolamine, the phosphatidylglycerol, the lipopolysaccharide, the lipoteichoic acid, and the cardiolipin, and is generally captured by the positively charged bacteriocins. The cationic charged groups of the bacteriocins electrostatically interact with the anionic bacterial cell surface, while the hydrophobic surfaces are attached to the membrane and traverse the lipid bilayer. The bacteriocins self-associate or polymerize to develop complexes after passing through the lipid bilayer. The bacteriocins induce the cell death by increasing the permeability of the bacterial membrane, forming pores that cause dissipation of the proton motive force, exhaustion of ATP, and leakage of intracellular substrates. The Gram-positive bacteria origin bacteriocins only perform for the Gram-positive bacteria and are not effective against the Gram-negative bacteria because of their different membrane compositions and the selective membrane permeability. The disadvantages could be compensated by mixing processing with other preservatives and the application of further preservation methods (209,210,211,212 and 213).

The Nisin

The Nisin is the most representative class I bacteriocin. The Nisin is produced by several strains of the Lactococcus lactis, a species that is widely used for the dairy production. The Nisin was first approved as a food preservative in the United Kingdom in the 1950s and is now widely used worldwide and is permitted in over 50 countries. The structure of the nisin consists of a polypeptide with 34 amino acids, a 3.5 kDa molecular mass, and contains the methyllanthionine and the lanthionine groups. The Nisin has antimicrobial activities against a wide range of the Grampositive bacteria, including the Staphylococcus spp., the Bacillus spp., the Listeria spp., and the Enterococcus spp. The Nisaplin is a typical commercial nisin formulation. The Nisin could provide long lasting bacteriostatic effects on the pathogenic bacteria in the beef jerky at room temperature. The shelf-life extensive effect of the nisin in the B. cereus inoculated with the beef jerky. The beef jerky without the nisin, the counts of the mesophilic bacteria and the B. cereus increasing is unlikely for the beef jerky treated with the nisin at 25 °C for 60 days. The B. cereus grew after three days in the 100 IU nisin/gram. The treated groups and after 21 days in the 500 IU/ gram nisin treated groups. The nisin containing fermentate from the L. lactis 537 strain was evaluated for the inhibition of the Listeria monocytogenes in ready to eat sliced ham. The addition of the fermentate to the ready to eat sliced ham led to an immediate decrease in the Listeria monocytogenes counts from three log Colony Forming Unit / gram to below the detection limit stored at 4 °C (20 Colony Forming Unit / gram). The Nisin with the cinnamaldehyde and the grapefruit seed extract presented synergistic antibacterial effects. It reduced the counts of the Listeria monocytogenes by three log Colony Forming Unit / gram in the raw pork loin at 4 °C for 12 hours. The minimum inhibitory concentration of the nisin against the Listeria monocytogenes was 250 ppm in the laboratory media, but it was possible to reduce the concentration of 5-6 ppm against the growth of Listeria monocytogenes by mixing with the natural antibacterial substances in the pork (214,215,216,217,218 and 219).

The Pediocin

The Pediococcus spp., the Pediococcus acidilactici, and the Pediococcus pentosaceus are the main pediocin producing strains. Pediocin was classified into the bacteriocin group class IIa, characterized as small non modified peptides (<5 kDa) comprising less than 50 amino acids. The

Clinical Oncology Case Reports Page 7 of 15

Remarkably, pediocin showed antimicrobial activity even at nanomolar concentrations. The Food grade pediocin containing formulations are commercially available and marketed as ALTA 2341 and MicroGARD. The Pediocin has been studied for the inhibition of the Listeria spp. for the meat preservation. The antibacterial activities of pediocin PA 1 in the frankfurters and the P. acidilactici MCH14, the pediocin PA 1 producing strain, in the Spanish dry fermented sausages were assessed against the Listeria monocytogenes and the Clostridium perfringens. Inside the frankfurters treated with 5000 bacteriocin units /mL of the pediocin PA 1 produced by the P. acidilactici the MCH14, the Listeria monocytogenes was reduced by 2 and 0.6 log Colony Forming Unit / gram after the storage at 4 °C for 60 days and at 15 °C for one month, respectively. The Clostridium perfringens decreased with 5000 BU/mL of the pediocin PA 1 by two and 0.8 log Colony Forming Unit / gram after the storage at 10 °C for 60 d and at 15 °C for one month, respectively. The growth of the Listeria monocytogenes was inhibited by the pediocin producing strain, the P. acidilactici MCH14, in the Spanish dry fermented sausages at two log Colony Forming Unit / gram compared to the control. The bacHA 6111-2, the pediocin from the P. acidilactici HA 6111-2, was applied to the Portuguese fermented meat sausage (the Alheira) with the HPP treatment (300 MPa, five min, 10 $^{\circ}$ C) to inhibit the Listeria innocua . The bacteriostatic effect was verified for high inoculation counts of the L. innocua at 4 °C for 60 days. The decreasing the inoculated L. innocua, antibacterial effect was observed below two log Colony Forming Unit / gram from day three of the storage until the end of the storage. The antibacterial activities of a mixed preparation containing the pediocin from the Pediococcus pentosaceus and the Murraya koenigii (the curry tree) berries in a raw goat meat emulsion at 4 °C for 9 days. The L. innocua was reduced for 4.1 log Colony Forming Unit / gram in the treated samples concentrations at 8.3 mL pediocin/1000 grams of the meat emulsion with 10% (v/w) Murraya koenigii berries extract at the end of the storage. The total viable count and the psychrophilic count were also observed decrease in the treated samples, 2.2 log Colony Forming Unit /gram and 1.6 log Colony Forming Unit / gram, respectively (220,221,222,223,224 and 225).

The Sakacin

The Sakacins, a class II bacteriocin, are mainly produced by the Lactobacillus sakei or the Lactobacillus curvatus strains. The Commercial sakacin products are currently not presented. Compared to the nisin and the pediocin, the sakacins have a relatively narrow antimicrobial spectrum, especially with effective inhibition against the Listeria species. The antibacterial effect of the sakacin producing strain, the L. sakei CWBI B1365, and the L. curvatus CWBI B28, on the fate of the Listeria monocytogenes in the raw beef and poultry. In the refrigerated (5 °C) the raw beef, the L. sakei induced a decrease in the Listeria monocytogenes concentration by 1.5 log Colony Forming Unit / gram after seven days to two log Colony Forming Unit / gram after 14 days, and below the detection limit at 21 days. The addition of the L. curvatus reduced the Listeria monocytogenes to below the detection limit after seven days. However, in the poultry, the bacteriocin producing strain did not affect the inhibition of the Listeria monocytogenes. It was assumed that the type of the meat may have influenced the bacteriocin production by the Lactic acid bacteria. The antibacterial activity of different bacteriocin preparations using the sakacin Q produced by the L. curvatus ACU 1 on the meat surface was evaluated against the L. innocua. The freeze-dried reconstituted cell free supernatant (3200 AU/mL) was effective for the inhibition of L. innocua on the meat surface, decreasing its bacterial cell number to the detection limit (<2 log Colony Forming Unit / gram) after two weeks of the storage at 4-5 °C. The adsorption of the sakacin Q to the meat products, the main ingredients, the meat proteins, and the fat tissues did not affect its antibacterial activity (226,227,228,229,230 and 231).

The Bacteriocin Like Inhibitory Substance

The Bacteriocin Like Inhibitory Substance are among the antimicrobial substances produced by the bacteria and are not completely categorized in terms of amino acid composition, molecular size, and nucleotide sequence. Inside the ready to eat pork ham, the antibacterial effects of the Bacteriocin Like Inhibitory Substance produced by the Pediococcus pentosaceus American Type Culture Collection 43200 were assessed and compared

with those of the commercially available nisin preparations (the Nisaplin). The Bacteriocin Like Inhibitory Substance showed effective antibacterial activity against the Listeria seeligeri by 0.74 log Colony Forming Unit / gram in the ready to eat ham stored at 4 °C after two days. The slight increase in the Listeria seeligeri counts was detected in the Bacteriocin Like Inhibitory Substance treated samples from six days to the end of the storage. The Nisaplin did not present any antibacterial effect for up to two days. After two days, the Nisaplin started to induce a decrease in the Listeria seeligeri counts throughout the refrigerated storage. This might have been due to the higher sensitivity of the Bacteriocin Like Inhibitory Substance to residual proteases compared to the nisin, thus weakening its antibacterial effect. The Bacteriocin Like Inhibitory Substance producing the Lactic acid bacteria strains, the P. acidilactici KTU05 7, the Pediococcus pentosaceus KTU05 9, and the Listeria sakei KTU05 6, were used to ferment the plant (the Jerusalem artichoke, the Helianthus tuberosus L.), and 5% of the fermented products were tested to inhibit the foodborne pathogen at 18 °C for half day in the ready to cook minced pork. The P. acidilactici fermented product presented the highest antimicrobial activity compared to the other strains. The counts of the Escherichia coli, the Enterococcus faecalis, the Staphylococcus aureus, and the Streptococcus spp. were reduced by 5.53, 4.37, 4.86, and 3.84 log Colony Forming Unit / gram, respectively, compared to the control groups, suggesting that the fermented product of the Bacteriocin Like Inhibitory Substance producing strains showed an enhanced antibacterial effect. The Bacteriocin Like Inhibitory Substance obtained from the Enterococcus faecium DB1 inhibited the growth and formation of the biofilms of the Clostridium perfringens in the chicken meat. The 2.5 mg/mL of DB1 Bacteriocin Like Inhibitory Substance suppressed the growth of the Clostridium perfringens by 30%. The Clostridium perfringens growth was inhibited by 50% at 5 mg/mL The DB1 Bacteriocin Like Inhibitory Substance. The Biofilm formation by the Clostridium perfringens treated with 5 mg/mL DB1 Bacteriocin Like Inhibitory Substance was radically reduced by 90% at 4 °C for three days compared to the control groups. The 2.5 mg/mL of the DB1 Bacteriocin Like Inhibitory Substance also inhibited biofilm formation by the Clostridium perfringens under the same conditions. The Bacteriocin Like Inhibitory Substance could inhibit the formation of the Clostridium perfringens biofilms on the chicken surfaces due to its antibacterial effect (232,233,234,235,236 and 237).

The Other Microorganism Sources

The mytichitin CB peptide, which was isolated from the blood lymphocytes of the Mytilus coruscus, showed antibacterial effects against the Gram-positive bacteria and the fungi. The mytichitin CB peptide expressed by Pichia pastorisi and applied it to the pork preservation. The total viable counts of the treated group with 6 mg/L of mytichitin CB derived from the P. pastorisi was reduced by 33% (1-2 log Colony Forming Unit / gram) compared to the control group after the storage at 4 °C for 5 days. The Mytichitin CB effectively inhibited the total bacterial growth during the storage compared to the groups treated with 50 mg/L of the nisin. The Mytichitin CB at 6 and 12 mg/L suppressed Staphylococcus spp. and Escherichia spp., respectively, with a reduction of 1–2 log Colony Forming Unit / gram, respectively. The Listeria spp. and the Pseudomonas spp. we're not detected during the storage, unlike the control and nisin treated groups. The Hispidalin is a unique Antimicrobial Peptides derived from the seeds of the Benincasa hispida and has been shown to exhibit the antimicrobial effects against the various bacteria. The hispidalin expressed by the P. pastorisi was used as a preservative for the pork. The Pork treated with 100 μg/mL hispidalin showed bacteriostatic effects during the entire refrigerated storage period. The total viable count of the pork with 100 μg/mL hispidalin was one log Colony Forming Unit /gram decrease than that of the control group at 4 °C for seven days (236,237, 238,239 and 240).

Conclusions

The Meat and its products are excellent nutrient sources due to their abundant protein content, the essential amino acids, the vitamins, and the minerals. The meat and its products are susceptible to the contamination by the foodborne pathogenic and the various spoilage bacteria because of their high-water activity and the nutrient content. The application of the

Clinical Oncology Case Reports Page 8 of 15

preservatives is an indispensable element in the livestock food processing to prevent the food poisoning, delay the spoilage, and extend their shelf life. The Industrial preservatives, commonly made up of the chemicals, are not demanded by the food customers because of their negative health concerns. The natural preservatives derived from the plants (the rosemary, the sage, the chestnut, the Grapefruit Seed Extract, and the tumeric), the animals (the lysozyme, the lactoferrin, the lactoferoxidase, the ovotransferrin, and others), and the bacteria (the organic acids, the bacteriocins, and the Bacteriocin Like Inhibitory Substance) have been explored as alternatives to the chemical preservatives. The versatility of the natural preservatives compared to the chemical preservatives is limited due to the production cost, the standardization, the insufficient toxicity studies, and the negative sensory effects on the food. To compensate for these disadvantages, various applications have been studied for their synergistic effect with the other natural preservatives with reduced the application concentrations compared to single use, the application of the physical treatment (the gamma irradation, the high-pressure processing, and the drying), the encapsulation, and the possibility of the packaging materials. The various natural preservatives and the application methods to inhibit the growth of the foodborne pathogenic and the spoilage bacteria in the livestock foods. The Natural preservatives are expected to be in high demand due to the consumer and the industrial requests. Therefore, it is necessary to explore various applications of the existing natural preservatives, while continuously searching for the novel ones.

Conflicts of Interest

The author declares no conflicts of interest

References:

- Shaltout, F.A., Riad,E.M., and AbouElhassan, Asmaa, A(2017): prevalence Of Mycobacterium Tuberculosis In Imported cattle Offals And Its lymph Nodes. Veterinary Medical Journal -Giza (VMJG), 63(2): 115 – 122.
- Papagianni M. Ribosomally synthesized peptides with antimicrobial properties: Biosynthesis, structure, function, and applications. Biotechnol. Adv. 2003;21:465–499.
- 3. Shaltout, F.A., Riad,E.M., and Asmaa Abou-Elhassan (2017): Prevalence Of Mycobacterium Spp. In Cattle Meat And Offal's Slaughtered In And Out Abattoir. Egyptian Veterinary medical Association, 77(2): 407 420.
- Meng D.-M., Sun S.-N., Shi L.-Y., Cheng L., Fan Z.-C. Application of antimicrobial peptide mytichitin-CB in pork preservation during cold storage. Food Control. 2021;125:108041.
- Abd Elaziz, O., Fatin S. Hassanin, Fahim A. Shaltout and Othman A. Mohamed (2021): Prevalence of Some Foodborne Parasitic Affection in Slaughtered Animals in Loacal Egyptian Abottoir. Journal of Nutrition Food Science and Technology 2(3): 1-5.
- Del Olmo A., Calzada J., Nuñez M. Effect of lactoferrin and its derivatives, high hydrostatic pressure, and their combinations, on Escherichia coli O157:H7 and Pseudomonas fluorescens in chicken filets. Innov. Food Sci. Emerg. Technol. 2012;13:51–56.
- Abd Elaziz, O., Fatin, S Hassanin, Fahim, A Shaltout, Othman, A Mohamed (2021): Prevalence of some zoonotic parasitic affections in sheep carcasses in a local abattoir in Cairo, Egypt. Advances in Nutrition & Food Science 6(2): 6(2): 25-31.
- Nieto-Lozano J.C., Reguera-Useros J.I., Peláez-Martínez M.d.C., Sacristán-Pérez-Minayo G., Gutiérrez-Fernández Á.J., de la Torre A.H. The effect of the pediocin PA-1 produced by Pediococcus acidilactici against Listeria monocytogenes and Clostridium perfringens in Spanish dryfermented sausages and frankfurters. Food Control. 2010;21:679–685.
- Al Shorman,A.A.M. ;Shaltout,F.A. and hilat,N (1999):Detection of certain hormone residues in meat marketed in Jordan.Jordan University of Science and

- Technology, 1st International Conference on Sheep and goat Diseases and Productivity, 23-25 October, 1999.
- 10. Müller-Auffermann K., Grijalva F., Jacob F., Hutzler M. Nisin and its usage in breweries: A review and discussion. J. Inst. Brew. 2015;121:309–319.
- 11. Ebeed Saleh, Fahim Shaltout, Essam Abd Elaal (2021); Effect of some organic acids on microbial quality of dressed cattle carcasses in Damietta abattoirs, Egypt. Damanhour Journal of Veterinary Sciences 5(2): 17-20.
- 12. Cegielska-Radziejewska R., Szablewski T., Radziejewska-Kubzdela E., Tomczyk Ł., Biadała A., Leśnierowski G. The effect of modified lysozyme treatment on the microflora, physicochemical and sensory characteristics of pork packaged in preservative gas atmospheres. Coatings. 2021;11:488.
- 13. Edris A, Hassanin, F. S; Shaltout, F.A., Azza H Elbaba and Nairoz M Adel(2017): Microbiological Evaluation of Some Heat Treated Fish Products in Egyptian Markets.EC Nutrition 12.3 (2017): 124-132.
- 14. Elliot R.M., McLay J.C., Kennedy M.J., Simmonds R.S. Inhibition of foodborne bacteria by the lactoperoxidase system in a beef cube system. Int. J. Food Microbiol. 2004;91:73–81.
- Edris ,A., Hassan,M.A., Shaltout,F.A. and Elhosseiny , S(2013): Chemical evaluation of cattle and camel meat.BENHA VETERINARY MEDICAL JOURNAL, 24(2): 191-197 .
- Rao M.S., Chander R., Sharma A. Synergistic effect of chitooligosaccharides and lysozyme for meat preservation. LWT. 2008;41:1995–2001.
- Edris ,A.M., Hassan,M.A., Shaltout,F.A. and Elhosseiny , S(2012): Detection of E.coli and Salmonella organisms in cattle and camel meat. BENHA VETERINARY MEDICAL JOURNAL, 24(2): 198-204.
- Przybylski R., Firdaous L., Chataigne G., Dhulster P., Nedjar N. Production of an antimicrobial peptide derived from slaughterhouse by-product and its potential application on meat as preservative. Food Chem. 2016;211:306–313.
- Edris A.M.; Hemmat M. I., Shaltout F.A.; Elshater M.A., Eman F.M.I. (2012): STUDY ON INCIPIENT SPOILAGE OF CHILLED CHICKEN CUTS-UP. BENHA VETERINARY MEDICAL JOURNAL, VOL. 23, NO. 1, JUNE 2012: 81-86
- Lee J.S., Park S.W., Lee H.B., Kang S.S. Bacteriocin-like inhibitory substance (BLIS) activity of Enterococcus faecium DB1 against biofilm formation by Clostridium perfringens. Probiotics Antimicrob. Proteins. 2021;13:1452–1457.
- 21. Edris A.M.; Hemmat M.I.; Shaltout F.A.; Elshater M.A., Eman, F.M.I.(2012):CHEMICAL ANALYSIS OF CHICKEN MEAT WITH RELATION TO ITS QUALITY. BENHA VETERINARY MEDICAL JOURNAL, 23(1): 87-92
- Rolfe V., Mackonochie M., Mills S., McLennan E. Turmeric/curcumin and health outcomes: A meta-review of systematic reviews. Eur. J. Integr. Med. 2020:101252.
- 23. Edris, A.M.; Shaltout, F.A. and Abd Allah, A.M. (2005): Incidence of Bacillus cereus in some meat products and the effect of cooking on its survival. Zag. Vet. J.33 (2):118-124.
- Dong A., Malo A., Leong M., Ho V.T.T., Turner M.S. Control
 of Listeria monocytogenes on ready-to-eat ham and fresh cut
 iceberg lettuce using a nisin containing Lactococcus lactis
 fermentate. Food Control. 2021;119:107420.
- 25. Edris, A.M.; Shaltout, F.A. and Arab, W.S. (2005): Bacterial Evaluation of Quail Meat. Benha Vet. Med.J.16 (1):1-14.
- Shwaiki L.N., Lynch K.M., Arendt E.K. Future of antimicrobial peptides derived from plants in food application—A focus on synthetic peptides. Trends Food Sci. Technol. 2021;112:312–324.
- Edris, A.M.; Shaltout, F.A.; Salem, G.H. and El-Toukhy, E.I. (2011): Incidence and isolation of Salmonellae from some meat products. Benha University, Faculty of Veterinary Medicine, Fourth Scientific Conference 25-27th May

- 2011Veterinary Medicine and Food Safety) 172-179 benha , Egypt.
- 28. Jaspal M.H., Ijaz M., Haq H.A.u., Yar M.K., Asghar B., Manzoor A., Badar I.H., Ullah S., Islam M.S., Hussain J. Effect of oregano essential oil or lactic acid treatments combined with air and modified atmosphere packaging on the quality and storage properties of chicken breast meat. LWT. 2021;146:111459.
- Edris AA, Hassanin, F. S; Shaltout, F.A., Azza H Elbaba and Nairoz M Adel.(2017): Microbiological Evaluation of Some Heat Treated Fish Products in Egyptian Markets. EC Nutrition 12.3 (2017): 134-142.
- 30. Wang F.-S. Effect of antimicrobial proteins from porcine leukocytes on Staphylococcus aureus and Escherichia coli in comminuted meats. Meat Sci. 2003;65:615–621.
- Edris, A.M.; Shaltout, F.A.; Salem, G.H. and El-Toukhy, E.I. (2011): Plasmid profile analysis of Salmonellae isolated from some meat products. Benha University, Faculty of Veterinary Medicine, Fourth Scientific Conference 25-27th May 2011Veterinary Medicine and Food Safety)194-201 benha, Egypt.
- 32. Seol K.H., Lim D.G., Jang A., Jo C., Lee M. Antimicrobial effect of kappa-carrageenan-based edible film containing ovotransferrin in fresh chicken breast stored at 5 degrees C. Meat Sci. 2009;83:479–483.
- Ragab A, Abobakr M. Edris, Fahim A.E. Shaltout, Amani M. Salem(2022): Effect of titanium dioxide nanoparticles and thyme essential oil on the quality of the chicken fillet. BENHA VETERINARY MEDICAL JOURNAL41(2): 38-40.
- Anastasio A., Marrone R., Chirollo C., Smaldone G., Attouchi M., Adamo P., Sadok S., Pepe T. Swordfish steaks vacuumpacked with Rosmarinus officinalis. Ital. J. Food Sci. 2014;26:390–397.
- 35. Woraprayote W., Malila Y., Sorapukdee S., Swetwiwathana A., Benjakul S., Visessanguan W. Bacteriocins from lactic acid bacteria and their applications in meat and meat products. Meat Sci. 2016;120:118–132.
- 36. Hassan, M.A, Shaltout, F. A, Arfa M.M, Mansour A.H and Saudi, K. R(2013): BIOCHEMICAL STUDIES ON RABBIT MEAT RELATED TO SOME DISEASES. BENHA VETERINARY MEDICAL JOURNAL 25(1):88-93.
- Mastromatteo M., Lucera A., Sinigaglia M., Corbo M.R. Synergic antimicrobial activity of lysozyme, nisin, and EDTA against Listeria monocytogenes in ostrich meat patties. J. Food Sci. 2010;75:M422–M429.
- 38. Hassan, M.A and Shaltout, F.A. (1997): Occurrence of Some Food Poisoning Microorganisms In Rabbit Carcasses Alex.J.Vet.Science, 13(1):55-61.
- 39. Lee N.-K., Paik H.-D. Prophylactic effects of probiotics on respiratory viruses including COVID-19: A review. Food Sci. Biotechnol. 2021;30:773–781.
- 40. Hassan M, Shaltout FA* and Saqur N (2020): Histamine in Some Fish Products. Archives of Animal Husbandry & Dairy Science 2(1): 1-3.
- 41. Wu T., Wu C., Fu S., Wang L., Yuan C., Chen S., Hu Y. Integration of lysozyme into chitosan nanoparticles for improving antibacterial activity. Carbohydr. Polym. 2017;155:192–200.
- 42. Hassan, M.A and Shaltout, F.A. (2004): Comparative Study on Storage Stability of Beef, Chicken meat, and Fish at Chilling Temperature. Alex.J.Vet.Science, 20(21):21-30.
- 43. Stimbirys A., Bartkiene E., Siugzdaite J., Augeniene D., Vidmantiene D., Juodeikiene G., Maruska A., Stankevicius M., Cizeikiene D. Safety and quality parameters of ready-to-cook minced pork meat products supplemented with Helianthus tuberosus L. tubers fermented by BLIS producing lactic acid bacteria. J. Food Sci. Technol. 2015;52:4306–4314.
- 44. Hassan, M.A; Shaltout, F.A.; Arafa, M.M.; Mansour, A.H. and Saudi, K.R.(2013): Biochemical studies on rabbit meat related to some diseases. Benha Vet. Med.J.25 (1):88-93.

- Lee D., Heinz V., Knorr D. Effects of combination treatments of nisin and high-intensity ultrasound with high pressure on the microbial inactivation in liquid whole egg. Innov. Food Sci. Emerg. Technol. 2003;4:387–393.
- Hassan, M.A; Shaltout, F.A.; Maarouf, A.A. and El-Shafey, W.S.(2014): Psychrotrophic bacteria in frozen fish with special reference to pseudomonas species .Benha Vet. Med.J.27 (1):78-83.
- 47. Lee N.K., Kim H.W., Lee J.Y., Ahn D.U., Kim C.J., Paik H.D. Antimicrobial effect of nisin against Bacillus cereus in beef jerky during storage. Korean J. Food Sci. Anim. Resour. 2015;35:272–276.
- Hassan, M.A; Shaltout, F.A.; Arafa, M.M.; Mansour, A.H. and Saudi, K.R.(2013): Bacteriological studies on rabbit meat related to some diseases
 Benha Vet. Med.J.25 (1):94-99.
- Yousefi M., Farshidi M., Ehsani A. Effects of lactoperoxidase system-alginate coating on chemical, microbial, and sensory properties of chicken breast fillets during cold storage. J. Food Saf. 2018;38:e12449.
- Hassanin, F. S; Hassan, M.A., Shaltout, F.A., Nahla A. Shawqy and 2Ghada A. Abd-Elhameed (2017): Chemical criteria of chicken meat. BENHA VETERINARY MEDICAL JOURNAL, 33(2):457-464.
- 51. Delves-Broughton J. Natural antimicrobials as additives and ingredients for the preservation of foods and beverages. In: Baines D., Seal R., editors. Natural Food Additives, Ingredients and Flavourings. 1st ed. Woodhead Publishing Series in Food Science, Technology and Nutrition; Cambridge, UK: 2012. pp. 127–161.
- 52. Hassanin, F. S; Hassan,M.A.; Shaltout, F.A. and Elrais-Amina, M(2014): CLOSTRIDIUM PERFRINGENS IN VACUUM PACKAGED MEAT PRODUCTS. BENHA VETERINARY MEDICAL JOURNAL, 26(1):49-53.
- 53. Kumar Y., Kaur K., Shahi A.K., Kairam N., Tyagi S.K. Antilisterial, antimicrobial and antioxidant effects of pediocin and Murraya koenigii berry extract in refrigerated goat meat emulsion. LWT. 2017;79:135–144.
- Hassanien, F.S.; Shaltout, F.A.; Fahmey, M.Z. and Elsukkary, H.F.(2020): Bacteriological quality guides in local and imported beef and their relation to public health. Benha Veterinary Medical Journal 39: 125-129.
- Montville T.J., Bruno M.E.C. Evidence that dissipation of proton motive force is a common mechanism of action for bacteriocins and other antimicrobial proteins. Int. J. Food Microbiol. 1994;24:53–74.
- Hassanin, F. S; Shaltout, F.A. and , Mostafa E.M(2013):
 Parasitic affections in edible offal. Benha Vet. Med.J.25 (2):34-39.
- Kotra V.S.R., Satyabanta L., Goswami T.K. A critical review of analytical methods for determination of curcuminoids in turmeric. J. Food Sci. Technol. 2019;56:5153–5166.
- Hassanin, F. S; Shaltout, F.A., Lamada, H.M., Abd Allah, E.M.(2011): THE EFFECT OF PRESERVATIVE (NISIN) ON THE SURVIVAL OF LISTERIA MONOCYTOGENES. BENHA VETERINARY MEDICAL JOURNAL (2011)-SPECIAL ISSUE [I]: 141-145.
- Ma B., Guo Y., Fu X., Jin Y. Identification and antimicrobial mechanisms of a novel peptide derived from egg white ovotransferrin hydrolysates. LWT. 2020;131:109720.
- Khattab, E., Fahim Shaltout and Islam Sabik (2021): Hepatitis A virus related to foods. BENHA VETERINARY MEDICAL JOURNAL 40(1): 174-179..
- 61. Sedighi R., Zhao Y., Yerke A., Sang S. Preventive and protective properties of rosemary (Rosmarinus officinalis L.) in obesity and diabetes mellitus of metabolic disorders: A brief review. Curr. Opin. Food Sci. 2015;2:58–70.
- 62. Saad M. Saad , Fahim A. Shaltout , Amal A. A. Farag & Hashim F. Mohammed (2022): Organophosphorus Residues in Fish in Rural Areas. Journal of Progress in Engineering and Physical Science 1(1): 27-31..

- 63. Dortu C., Huch M., Holzapfel W.H., Franz C.M., Thonart P. Anti-listerial activity of bacteriocin-producing Lactobacillus curvatus CWBI-B28 and Lactobacillus sakei CWBI-B1365 on raw beef and poultry meat. Lett. Appl. Microbiol. 2008;47:581–586.
- 64. Saif, M., Saad S.M., Hassanin, F. S; Shaltout FA, Marionette Zaghloul (2019): Molecular detection of enterotoxigenic Staphylococcus aureus in ready-to-eat beef products. Benha Veterinary Medical Journal 37 (2019) 7-11.
- 65. Massantini R., Moscetti R., Frangipane M.T. Evaluating progress of chestnut quality: A review of recent developments. Trends Food Sci. Technol. 2021;113:245–254.
- Saif,M., Saad S.M., Hassanin, F. S; Shaltout, F.A., Marionette Zaghlou (2019); Prevalence of methicillinresistant Staphylococcus aureus in some ready-to-eat meat products. Benha Veterinary Medical Journal 37 (2019) 12-15.
- Papagianni M., Anastasiadou S. Pediocins: The bacteriocins of Pediococci. Sources, production, properties and applications. Microb. Cell Factories. 2009;8:3.
- Farag, A. A., Saad M. Saad¹, Fahim A. Shaltout1, Hashim F. Mohammed(2023 a): Studies on Pesticides Residues in Fish in Menofia Governorate. Benha Journal of Applied Sciences ,. 8(5): 323-330.
- Castro S., Silva J., Casquete R., Queirós R., Saraiva J., Teixeira P. Combined effect of pediocin bacHA-6111-2 and high hydrostatic pressure to control Listeria innocua in fermented meat sausage. Int. Food Res. J. 2018;25:553–560.
- Farag, A. A., Saad M. Saad¹, Fahim A. Shaltout1, Hashim F. Mohammed(2023 b): Organochlorine Residues in Fish in Rural Areas. Benha Journal of Applied Sciences, 8 (5): 331-336
- 71. Kumariya R., Garsa A.K., Rajput Y.S., Sood S.K., Akhtar N., Patel S. Bacteriocins: Classification, synthesis, mechanism of action and resistance development in food spoilage causing bacteria. Microb. Pathog. 2019;128:171–177.
- 72. Shaltout, F.A., Mona N. Hussein, Nada Kh. Elsayed (2023): Histological Detection of Unauthorized Herbal and Animal Contents in Some Meat Products. Journal of Advanced Veterinary Research 13(2): 157-160.
- Marchese A., Orhan I.E., Daglia M., Barbieri R., Di Lorenzo A., Nabavi S.F., Gortzi O., Izadi M., Nabavi S.M. Antibacterial and antifungal activities of thymol: A brief review of the literature. Food Chem. 2016;210:402–414.
- Shaltout, F. A., Heikal, G. I., Ghanem, A. M.(2022): Mycological quality of some chicken meat cuts in Gharbiya governorate with special reference to Aspergillus flavus virulent factors. benha veteriv medical journal veterinary 42(1): 12-16.
- 75. De Azevedo P.O.d.S., Converti A., Gierus M., de Souza Oliveira R.P. Antimicrobial activity of bacteriocin-like inhibitory substance produced by Pediococcus pentosaceus: From shake flasks to bioreactor. Mol. Biol. Rep. 2019;46:461–469.
- Shaltout, F.A., Ramadan M. Salem, Eman M. Eldiasty, Fatma A. Diab (2022): Seasonal Impact on the Prevalence of Yeast Contamination of Chicken Meat Products and Edible Giblets. Journal of Advanced Veterinary Research 12(5): 641-644.
- 77. Shahnawaz M., Soto C. Microcin amyloid fibrils A are reservoir of toxic oligomeric species. J. Biol. Chem. 2012;287:11665–11676.
- 78. Shaltout, F.A., Abdelazez Ahmed Helmy Barr and Mohamed Elsayed Abdelaziz (2022): Pathogenic Microorganisms in Meat Products. Biomedical Journal of Scientific & Technical Research 41(4): 32836-32843.
- 79. De Alba M., Bravo D., Medina M. Inactivation of Listeria monocytogenes and Salmonella Enteritidis in dry-cured ham by combined treatments of high pressure and the lactoperoxidase system or lactoferrin. Innov. Food Sci. Emerg. Technol. 2015;31:54–59.

 Shaltout, F.A., Thabet, M.G. and Koura, H.A. (2017). Impact of Some Essential Oils on the Quality Aspect and Shelf Life of Meat. J Nutr Food Sci., 7: 647.

Page 10 of 15

- 81. Moon S.H., Paik H.D., White S., Daraba A., Mendonca A.F., Ahn D.U. Influence of nisin and selected meat additives on the antimicrobial effect of ovotransferrin against Listeria monocytogenes. Poult. Sci. 2011;90:2584–2591.
- Shaltout, F.A.,., Islam Z. Mohammed², El -Sayed A. Afify(2020): Bacteriological profile of some raw chicken meat cuts in Ismailia city, Egypt.Benha Veterinary Medical Journal 39 (2020) 11-15.
- 83. Ko K.Y., Mendonca A.F., Ahn D.U. Influence of zinc, sodium bicarbonate, and citric acid on the antibacterial activity of ovotransferrin against Escherichia coli O157:H7 and Listeria monocytogenes in model systems and ham. Poult. Sci. 2008;87:2660–2670.
- 84. Shaltout, F.A., Islam, Z. Mohammed²., El -Sayed A. Afify(2020): Detection of E. coli O157 and Salmonella species in some raw chicken meat cuts in Ismailia province, Egypt. Benha Veterinary Medical Journal 39 (2020) 101-104.
- 85. Meng D.-M., Sun X.-Q., Sun S.-N., Li W.-J., Lv Y.-J., Fan Z.-C. The potential of antimicrobial peptide Hispidalin application in pork preservation during cold storage. J. Food Process. Preserv. 2020;44:e14443.
- 86. Shaltout, F.A., E.M. El-diasty and M. A. Asmaa- Hassan (2020): HYGIENIC QUALITY OF READY TO EAT COOKED MEAT IN RESTAURANTS AT Cairo. Journal of Global Biosciences 8(12): 6627-6641..
- 87. Giansanti F., Panella G., Leboffe L., Antonini G. Lactoferrin from milk: Nutraceutical and pharmacological properties. Pharmaceuticals. 2016;9:61.
- Shaltout, F.A., Marrionet Z. Nasief, L. M. Lotfy, Bossi T. Gamil(2019): Microbiological status of chicken cuts and its products. Benha Veterinary Medical Journal 37 (2019) 57-63.
- Vasconcelos N., Croda J., Simionatto S. Antibacterial mechanisms of cinnamon and its constituents: A review. Microb. Pathog. 2018;120:198–203.
- Shaltout, F.A.(2019): Poultry Meat. Scholarly Journal of Food and Nutrition 22 1-2..
- 91. Mei J., Ma X., Xie J. Review on natural preservatives for extending fish shelf life. Foods. 2019;8:490.
- 92. Shaltout, F.A.(2019): Food Hygiene and Control. Food Science and Nutrition Technology 4(5): 1-2.
- 93. Yuan S., Yin J., Jiang W., Liang B., Pehkonen S., Choong C. Enhancing antibacterial activity of surface-grafted chitosan with immobilized lysozyme on bioinspired stainless steel substrates. Colloids Surf. B. 2013;106:11–21.
- Hassanin, F. S; Shaltout, F.A., Seham N. Homouda and Safaa M. Arakeeb(2019): Natural preservatives in raw chicken meat. Benha Veterinary Medical Journal 37 (2019) 41-45.
- 95. Xu M.M., Kaur M., Pillidge C.J., Torley P.J. Microbial biopreservatives for controlling the spoilage of beef and lamb meat: Their application and effects on meat quality. Crit. Rev. Food Sci. Nutr. 2021:1–35.
- 96. Hazaa, W., Shaltout, F.A., Mohamed El-Shate(2019): Prevalence of some chemical hazards in some meat products. Benha Veterinary Medical Journal 37 (2) 32-36.
- Rivas F.P., Castro M.P., Vallejo M., Marguet E., Campos C.A. Sakacin Q produced by Lactobacillus curvatus ACU-1: Functionality characterization and antilisterial activity on cooked meat surface. Meat Sci. 2014;97:475

 –479.
- 98. Hazaa, W, Shaltout, F.A., Mohamed El-Shater (2019): Identification of Some Biological Hazards in Some Meat Products. Benha Veterinary Medical Journal 37 (2) 27-31.
- De Azevedo P.O.S., Mendonca C.M.N., Seibert L., Dominguez J.M., Converti A., Gierus M., Oliveira R.P.S. Bacteriocin-like inhibitory substance of Pediococcus pentosaceus as a biopreservative for Listeria sp. control in ready-to-eat pork ham. Braz. J. Microbiol. 2020;51:949–956.

100. Gaafar,R. , Hassanin, F. S; Shaltout, F.A., Marionette Zaghloul (2019): Molecular detection of enterotoxigenic Staphylococcus aureus in some ready to eat meat-based sandwiches. Benha Veterinary Medical Journal 37 (2) 22-26.

- 101. Parada J.L., Caron C.R., Medeiros A.B.P., Soccol C.R. Bacteriocins from lactic acid bacteria: Purification, properties and use as biopreservatives. Braz. Arch. Biol. Technol. 2007;50:512–542.
- 102. Gaafar, R., Hassanin, F. S; Shaltout, F.A., Marionette Zaghloul(2019): Hygienic profile of some ready to eat meat product sandwiches sold in Benha city, Qalubiya Governorate, Egypt. Benha Veterinary Medical Journal 37 (2) 16-21.
- 103. Galvez A., Abriouel H., Benomar N., Lucas R. Microbial antagonists to food-borne pathogens and biocontrol. Curr. Opin. Biotechnol. 2010;21:142–148.
- 104. Saad S.M., Shaltout, F.A., Nahla A Abou Elroos, Saber B Elnahas(2019): Antimicrobial Effect of Some Essential Oils on Some Pathogenic Bacteria in Minced Meat. J Food Sci Nutr Res. 2019; 2 (1): 012-020.
- 105. Yu H.H., Song M.W., Song Y.J., Lee N.K., Paik H.D. Antibacterial effect of a mixed natural preservative against Listeria monocytogenes on lettuce and raw pork loin. J. Food Prot. 2019;82:2001–2006.
- 106. Tosati J.V., Messias V.C., Carvalho P.I.N., Rodrigues Pollonio M.A., Meireles M.A.A., Monteiro A.R. Antimicrobial effect of edible coating blend based on turmeric starch residue and gelatin applied onto fresh frankfurter sausage. Food Bioproc. Technol. 2017;10:2165–2175.
- 107. Saad S.M., Hassanin, F. S.; Shaltout, F.A., Marionette Z Nassif, Marwa Z Seif.(2019: Prevalence of Methicillin-Resistant Staphylococcus Aureus in Some Ready-to-Eat Meat Products. American Journal of Biomedical Science & Research 4(6):460-464.
- 108. Gogliettino M., Balestrieri M., Ambrosio R.L., Anastasio A., Smaldone G., Proroga Y.T., Moretta R., Rea I., De Stefano L., Agrillo B. Extending the shelf-life of meat and dairy products via PET-modified packaging activated with the antimicrobial peptide MTP1. Front. Microbiol. 2020;10:2963.
- 109. Shaltout, Fahim(2019): Pollution of Chicken Meat and Its Products by Heavy Metals. Research and Reviews on Healthcare: Open Access Journal, 4, 3(381-3382).
- 110. Chen X., Chen W., Lu X., Mao Y., Luo X., Liu G., Zhu L., Zhang Y. Effect of chitosan coating incorporated with oregano or cinnamon essential oil on the bacterial diversity and shelf life of roast duck in modified atmosphere packaging. Food Res. Int. 2021;147:110491.
- 111. Shaltout, F. A.; E.M EL-diasty; M. S. M Mohamed (2018): Effects of chitosan on quality attributes fresh meat slices stored at 4 C. BENHA VETERINARY MEDICAL JOURNAL, VOL. 35, NO. 2: 157-168.
- 112. Lazzaro B.P., Zasloff M., Rolff J. Antimicrobial peptides: Application informed by evolution. Science. 2020;368:6490.
- 113. Shaltout, F. A. and Abdel-Aziz, 2004: Salmonella enterica serovar Enteritidis in poultry meat and their epidemiology. Vet. Med. J. Giza, 52 (2004), pp. 429-436.
- 114. Borrajo P., Pateiro M., Barba F.J., Mora L., Franco D., Toldrá F., Lorenzo J.M. Antioxidant and antimicrobial activity of peptides extracted from meat by-products: A review. Food Anal. Methods. 2019;12:2401–2415.
- 115. Shaltout, F.A., Hala F El-Shorah, Dina I El Zahaby, Lamiaa M Lotfy(2018):Bacteriological Profile of Chicken Meat Products. SciFed Food & Dairy Technology Journal, 2:3.
- 116. Cegielka A., Hac-Szymanczuk E., Piwowarek K., Dasiewicz K., Slowinski M., Wronska K. The use of bioactive properties of sage preparations to improve the storage stability of low-pressure mechanically separated meat from chickens. Poult. Sci. 2019;98:5045–5053.
- 117. Eslamloo K., Falahatkar B., Yokoyama S. Effects of dietary bovine lactoferrin on growth, physiological performance, iron metabolism and non-specific immune responses of Siberian

- sturgeon Acipenser baeri. Fish Shellfish Immunol. 2012;32:976–985.
- 118. Shaltout, F.A., Mohamed, A.H. El-Shater., Wafaa Mohamed Abd El-Aziz(2015): Bacteriological assessment of Street Vended Meat Products sandwiches in kalyobia Governorate. BENHA VETERINARY MEDICAL JOURNAL, 28(2:)58-66
- 119. El-Saadony M.T., Abd El-Hack M.E., Swelum A.A., Al-Sultan S.I., El-Ghareeb W.R., Hussein E.O.S., Ba-Awadh H.A., Akl B.A., Nader M.M. Enhancing quality and safety of raw buffalo meat using the bioactive peptides of pea and red kidney bean under refrigeration conditions. Ital. J. Anim. Sci. 2021;20:762–776.
- 120. Shaltout, F.A., Mohamed A El shatter and Heba M Fahim(2019): Studies on Antibiotic Residues in Beef and Effect of Cooking and Freezing on Antibiotic Residues Beef Samples. Scholarly Journal of Food and Nutritionm 2(1) 1-4
- 121. Heymich M.L., Srirangan S., Pischetsrieder M. Stability and activity of the antimicrobial peptide Leg1 in solution and on meat and its optimized generation from chickpea storage protein. Foods. 2021;10:1192.
- 122. Shaltout FA, Zakaria IM and Nabil ME. (2018): Incidence of Some Anaerobic Bacteria Isolated from Chicken Meat Products with Special Reference to Clostridium perfringens. Nutrition and Food Toxicology 2.5 (2018): 429-438.
- 123. Khaleque M.A., Keya C.A., Hasan K.N., Hoque M.M., Inatsu Y., Bari M.L. Use of cloves and cinnamon essential oil to inactivate Listeria monocytogenes in ground beef at freezing and refrigeration temperatures. LWT. 2016;74:219–223.
- 124. Shaltout FA, Ahmed A A Maarouf and Mahmoud ES Elkhouly. (2017): Bacteriological Evaluation of Frozen Sausage. Nutrition and Food Toxicology 1.5; 174-185.
- 125. Stojanović-Radić Z., Pejčić M., Joković N., Jokanović M., Ivić M., Šojić B., Škaljac S., Stojanović P., Mihajilov-Krstev T. Inhibition of Salmonella Enteritidis growth and storage stability in chicken meat treated with basil and rosemary essential oils alone or in combination. Food Control. 2018;90:332–343.
- 126. Shaltout FA, El-Toukhy EI and Abd El-Hai MM.(2019): Molecular Diagnosis of Salmonellae in Frozen Meat and Some Meat Products. Nutrition and Food Technology Open Access 5(1): 1-6.
- 127. Moura-Alves M., Gouveia A.R., de Almeida J.M.M.M., Monteiro-Silva F., Silva J.A., Saraiva C. Behavior of Listeria monocytogenes in beef Sous vide cooking with Salvia officinalis L. essential oil, during storage at different temperatures. LWT. 2020;132:109896.
- 128. Shaltout, F.A., A.M.Ali and S.M.Rashad (2016): Bacterial Contamination of Fast Foods. Benha Journal of Applied Sciences (BJAS) 1 (2)45-51.
- 129. Yu H.H., Kim Y.J., Park Y.J., Shin D.-M., Choi Y.-S., Lee N.-K., Paik H.-D. Application of mixed natural preservatives to improve the quality of vacuum skin packaged beef during refrigerated storage. Meat Sci. 2020;169:108219.
- 130. Shaltout, F.A., Zakaria. I. M., Jehan Eltanani, Asmaa. Elmelegy(2015): Microbiological status of meat and chicken received to University student hostel. BENHA VETERINARY MEDICAL JOURNAL, 29(2):187-192, DECEMBER, 2015.
- 131. Selmi S., Rtibi K., Grami D., Sebai H., Marzouki L. Rosemary (Rosmarinus officinalis) essential oil components exhibit antihyperglycemic, anti-hyperlipidemic and antioxidant effects in experimental diabetes. Pathophysiology. 2017;24:297–303.
- 132. Saad,S.M.;Edris, A.M.; Shaltout,F.A. and Edris, Shimaa(2012): Isolation and identification of salmonellae and E.coli from meat and poultry cuts by using A.multiplex PCR. Benha Vet. Med.J.special issue 16-26.
- 133. Arshad M.S., Amjad Z., Yasin M., Saeed F., Imran A., Sohaib M., Anjum F.M., Hussain S. Quality and stability evaluation

- of chicken meat treated with gamma irradiation and turmeric powder. Int. J. Food Prop. 2019;22:154–172.
- 134. Saad, S.M. and Shaltout, F.A.(1998):Mycological Evaluation of camel carcasses at Kalyobia Abattoirs. Vet.Med.J. Giza,46(3):223-229.
- 135. Smaoui S., Hlima H.B., Braïek O.B., Ennouri K., Mellouli L., Khaneghah A.M. Recent advancements in encapsulation of bioactive compounds as a promising technique for meat preservation. Meat Sci. 2021;181:108585.
- 136. Beya M.M., Netzel M.E., Sultanbawa Y., Smyth H., Hoffman L.C. Plant-based phenolic molecules as natural preservatives in comminuted meats: A review. Antioxidants. 2021;10:263.
- 137. Saad S.M., Hassanin, F. S; Shaltout, F.A., Marionette Z Nassif, Marwa Z Seif.(2019): Prevalence of Methicillin-Resistant Staphylococcus Aureus in Some Ready-to-Eat Meat Products. American Journal of Biomedical Science & Research 4(6):460-464.
- 138. Radunz M., Dos Santos Hackbart H.C., Camargo T.M., Nunes C.F.P., de Barros F.A.P., Dal Magro J., Filho P.J.S., Gandra E.A., Radunz A.L., da Rosa Zavareze E. Antimicrobial potential of spray drying encapsulated thyme (Thymus vulgaris) essential oil on the conservation of hamburger-like meat products. Int. J. Food Microbiol. 2020;330:108696.
- 139. Saad S.M., Shaltout, F.A., Nahla A Abou Elroos and Saber B El-nahas. (2019): Incidence of Staphylococci and E. coli in Meat and Some Meat Products. EC Nutrition 14.6 (2019).
- 140. Zamuz S., Lopez-Pedrouso M., Barba F.J., Lorenzo J.M., Dominguez H., Franco D. Application of hull, bur and leaf chestnut extracts on the shelf-life of beef patties stored under MAP: Evaluation of their impact on physicochemical properties, lipid oxidation, antioxidant, and antimicrobial potential. Food Res. Int. 2018;112:263–273.
- 141. Shaltout FA, Riad EM,TES Ahmed and AbouElhassan A.(2017): Studying the Effect of Gamma Irradiation on Bovine Offal's Infected with Mycobacterium tuberculosis Bovine Type. Journal of Food Biotechnology Research 1 (6): 1-5.
- 142. Kiprotich S., Mendonca A., Dickson J., Shaw A., Thomas-Popo E., White S., Moutiq R., Ibrahim S.A. Thyme oil enhances the inactivation of Salmonella enterica on raw chicken breast meat during marination in lemon juice with added Yucca schidigera extract. Front. Nutr. 2020;7:619023.
- 143. Shaltout FA, Ahmed A A Maarouf and Mahmoud ES Elkhouly.(2017): Bacteriological Evaluation of Frozen Sausage. Nutrition and Food Toxicology 1.5 (2017): 174-185.
- 144. Lourenço T., Mendonça E., Nalevaiko P., Melo R., Silva P., Rossi D. Antimicrobial effect of turmeric (Curcuma longa) on chicken breast meat contamination. Braz. J. Poult. Sci. 2013;15:79–82.
- 145. Shaltout FA, Zakaria IM and Nabil ME.(2018): Incidence of Some Anaerobic Bacteria Isolated from Chicken Meat Products with Special Reference to Clostridium perfringens. Nutrition and Food Toxicology 2.5 (2018): 429-438.
- 146. Lee N.-K., Jung B.S., Na D.S., Yu H.H., Kim J.-S., Paik H.-D. The impact of antimicrobial effect of chestnut inner shell extracts against Campylobacter jejuni in chicken meat. LWT. 2016;65:746–750.
- 147. Shaltout FA, Mohamed, A.Hassan and Hassanin, F. S(2004): THERMAL INACTIVATION OF ENTEROHAEMORRHAGIC ESCHERICHIA COLI O157:H7 AND ITS SENSTIVITY TO NISIN AND LACTIC ACID CULTURES. 1rst Ann. Confr. , FVM., Moshtohor, Sept, 2004.
- 148. Hulankova R., Borilova G., Steinhauserova I. Combined antimicrobial effect of oregano essential oil and caprylic acid in minced beef. Meat Sci. 2013;95:190–194.
- 149. Shaltout FA, El-diasty, E,M. ;Elmesalamy, M. and Elshaer, M.(2014): Study on fungal contamination of some chicken meat products with special reference to 2 the use of PCR for

- its identification. Conference, Veterinary Medical Journal Giza vol. December 2014/12/17 vol.60: 1-10.
- 150. Hernandez H., Frankova A., Sykora T., Kloucek P., Kourimska L., Kucerova I., Banout J. The effect of oregano essential oil on microbial load and sensory attributes of dried meat. J. Sci. Food Agric. 2017;97:82–87.
- 151. Shaltout, F.A.(2002): Microbiological Aspects of Semicooked chicken Meat Products. Benha Veterinary Medical Journal 13.2.: 15-26.
- 152. Shange N., Makasi T., Gouws P., Hoffman L.C. Preservation of previously frozen black wildebeest meat (Connochaetes gnou) using oregano (Oreganum vulgare) essential oil. Meat Sci. 2019;148:88–95.
- 153. Lages L.Z., Radünz M., Gonçalves B.T., Silva da Rosa R., Fouchy M.V., de Cássia dos Santos da Conceição R., Gularte M.A., Barboza Mendonça C.R., Gandra E.A. Microbiological and sensory evaluation of meat sausage using thyme (Thymus vulgaris, L.) essential oil and powdered beet juice (Beta vulgaris L., Early Wonder cultivar) LWT. 2021;148:109896.
- 154. Shaltout FA, Mohammed Farouk; Hosam A.A. Ibrahim and Mostafa E.M. Afifi4.2017: Incidence of Coliform and Staphylococcus aureus in ready to eat fast foods. BENHA VETERINARY MEDICAL JOURNAL, 32(1): 13 - 17, MARCH, 2017.
- 155. De Oliveira T.L., de Araujo Soares R., Ramos E.M., das Gracas Cardoso M., Alves E., Piccoli R.H. Antimicrobial activity of Satureja montana L. essential oil against Clostridium perfringens type A inoculated in mortadella-type sausages formulated with different levels of sodium nitrite. Int. J. Food Microbiol. 2011;144:546–555.
- 156. Shaltout, F.A., Zakaria, I.M., Nabil, M.E.(2017): Detection and typing of Clostridium perfringens in some retail chicken meat products.BENHA VETERINARY MEDICAL JOURNAL., 33(2):283-291.
- 157. Zhu Y., Li C., Cui H., Lin L. Encapsulation strategies to enhance the antibacterial properties of essential oils in food system. Food Control. 2020;123:107856.
- 158. Shaltout, F.A.(1992): Studies on Mycotoxins in Meat and Meat by Products. M.V.Sc Thesis Faculty of Veterinary Medicine, Moshtohor, Zagazig University Benha branch.
- 159. Juneja V.K., Fan X., Peña-Ramos A., Diaz-Cinco M., Pacheco-Aguilar R. The effect of grapefruit extract and temperature abuse on growth of Clostridium perfringens from spore inocula in marinated, sous-vide chicken products. Innov. Food Sci. Emerg. Technol. 2006;7:100–106.
- 160. Shaltout, F.A.(1996): Mycological And Mycotoxicological profile Of Some Meat products. Ph.D.Thesis, Faculty of Veterinary Medicine, Moshtohor, Zagazig University Benha branch.
- 161. Maes C., Bouquillon S., Fauconnier M.-L. Encapsulation of essential oils for the development of biosourced pesticides with controlled release: A review. Molecules. 2019;24:2539.
- 162. Shaltout, F.A. (1998): Proteolytic Psychrotrophes in Some Meat products. Alex. Vet. Med. J.14 (2):97-107.
- 163. Ben-Fadhel Y., Cingolani M.C., Li L., Chazot G., Salmieri S., Horak C., Lacroix M. Effect of γ-irradiation and the use of combined treatments with edible bioactive coating on carrot preservation. Food Packag. Shelf Life. 2021;28:100635.
- 164. Shaltout, F.A.(1999): Anaerobic Bacteria in Vacuum Packed Meat Products. Benha Vet. Med.J.10 (1):1-10.
- 165. Soyer F., Keman D., Eroglu E., Ture H. Synergistic antimicrobial effects of activated lactoferrin and rosemary extract in vitro and potential application in meat storage. J. Food Sci. Technol. 2020;57:4395–4403.
- 166. Shaltout,F.A.(2000):Protozoal Foodborne Pathogens in some Meat Products. Assiut Vet. Med. J. 42 (84):54-59.
- 167. Kahraman T., Issa G., Bingol E.B., Kahraman B.B., Dumen E. Effect of rosemary essential oil and modified-atmosphere packaging (MAP) on meat quality and survival of pathogens in poultry fillets. Braz. J. Microbiol. 2015;46:591–599.

168. Shaltout,F.A.(2001): Quality evaluation of sheep carcasses slaughtered at Kalyobia abattoirs. Assiut Veterinary Medical Journal, 46(91):150-159.

- 169. Motavaf F., Mirvaghefi A., Farahmand H., Hosseini S.V. Effect of Zataria multiflora essential oil and potassium sorbate on inoculated Listeria monocytogenes, microbial and chemical quality of raw trout fillet during refrigerator storage. Food Sci. Nutr. 2021;9:3015–3025.
- 170. Shaltout, F.A.(2002): Microbiological Aspects of Semicooked Chicken Meat Products. Benha Vet.Med.J. 13(2):15-26
- 171. Sojic B., Pavlic B., Ikonic P., Tomovic V., Ikonic B., Zekovic Z., Kocic-Tanackov S., Jokanovic M., Skaljac S., Ivic M. Coriander essential oil as natural food additive improves quality and safety of cooked pork sausages with different nitrite levels. Meat Sci. 2019;157:107879.
- 172. Shaltout, F.A. (2003): Yersinia Enterocolitica in some meat products and fish marketed at Benha city. The Third international conference Mansoura 29-30 April.
- 173. Asioli D., Aschemann-Witzel J., Caputo V., Vecchio R., Annunziata A., Næs T., Varela P. Making sense of the "clean label" trends: A review of consumer food choice behavior and discussion of industry implications. Food Res. Int. 2017;99:58–71.
- 174. Shaltout, F.A.(2009):Microbiological quality of chicken carcasses at modern Poultry plant. The 3rd Scientific Conference, Faculty of Vet. Med., Benha University, 1-3 january.
- 175. Coutinho de Oliveira T.L., Malfitano de Carvalho S., de Araújo Soares R., Andrade M.A., Cardoso M.d.G., Ramos E.M., Piccoli R.H. Antioxidant effects of Satureja montana L. essential oil on TBARS and color of mortadella-type sausages formulated with different levels of sodium nitrite. LWT. 2012;45:204–212.
- 176. Shaltout,F.A. and Abdel Aziz ,A.M.(2004): Salmonella enterica Serovar Enteritidis in Poultry Meat and their Epidemiology .Vet.Med.J.,Giza,52(3):429-436.
- 177. World Health Organization . High-Dose Irradiation: Wholesomeness of Food Irradiatied with Doses above 10 kGy. World Health Organization; Geneva, Switzerland: 1999.
- 178. Shaltout,F.A. and Abdel Aziz ,A.M.(2004): ESCHERICHIA COLI STRAINS IN SLAUGHTERED ANIMALS AND THEIR PUBLIC HEALTH IMPORTENCE. J.Egypt. Vet. Med. Association 64(2):7-21.
- 179. Abdeldaiem M. Using of combined treatment between edible coatings containing ethanolic extract of papaya (carica papaya L.) leaves and gamma irradiation for extending shelf-life of minced chicken meat. Am. J. Food Technol. 2014;2:6–16.
- 180. Shaltout, F.A., Amin, R., Marionet, Z., Nassif and Shimaa, Abdel-wahab (2014): Detection of aflatoxins in some meat products. Benha veterinary medical journal, 27(2):368-374.
- 181. Akhter R., Masoodi F., Wani T.A., Rather S.A., Hussain P.R. Synergistic effect of low dose γ-irradiation, natural antimicrobial and antioxidant agents on quality of meat emulsions. Radiat. Phys. Chem. 2021;189:109724.
- 182. Shaltout,F.A. and Afify, Jehan Riad,EM and Abo Elhasan, Asmaa,A.(2012): Improvement of microbiological status of oriental sausage. Journal of Egyptian Veterinary Medical Association 72(2):157-167.
- 183. European Commission EU Guidance to the Commission Regulation (EC) No 450/2009 of 29 May 2009 on Active and Intelligent Materials and Articles Intended to Come into the Contact with Food. [(accessed on 12 October 2021)]. Available online: https://ec.europa.eu/food/safety/chemical_safety/food_contact_materials_en
- 184. Shaltout,F.A. and Daoud, J. R.(1996): Chemical analytical studies on rabbit meat and liver. Benha Vet. Med.J.8 (2):17-27.

- 185. Ming Y., Chen L., Khan A., Wang H., Wang C. Effects of tea polyphenols on physicochemical and antioxidative properties of whey protein coating. Food Sci. Biotechnol. 2020;29:1655– 1663
- 186. Shaltout,F.A. and Edris, A.M.(1999): Contamination of shawerma with pathogenic yeasts. Assiut Veterinary Medical Journal,40(64):34-39.
- 187. Balasubramaniam V., Martinez-Monteagudo S.I., Gupta R. Principles and application of high pressure—based technologies in the food industry. Annu. Rev. Food Sci. 2015;6:435–462.
- 188. Shaltout, F. A. ;Eldiasty, E. and Mohamed, M.S.(2014): Incidence of lipolytic and proteolytic fungi in some chicken meat products and their public health significance. Animal Health Research Institute: First International Conference on Food Safety and Technology 19-23 June 2014 Cairo Egypt pages 79-89.
- 189. Chuang S., Sheen S. High pressure processing of raw meat with essential oils-microbial survival, meat quality, and models: A review. Food Control. 2021;132:108529.
- 190. Shaltout, F.A.; Eldiasty, E.; Salem, R. and Hassan, Asmaa (2016): Mycological quality of chicken carcasses and extending shelf life by using preservatives at refrigerated storage. Veterinary Medical Journal -Giza (VMJG)62(3)1-7.
- 191. Pedreschi F., Mariotti-Celis M.S. Genetically Modified and Irradiated Food. Academic Press; Cambridge, MA, USA: 2020. Irradiation kills microbes: Can it do anything harmful to the food? pp. 233–242.
- 192. Shaltout, F.A.; Salem, R. Eldiasty, E.; and Diab, Fatema. (2016): Mycological evaluation of some ready to eat meat products with special reference to molecular chacterization. Veterinary Medical Journal -Giza 62(3)9-14.
- 193. Global Newswire The "Clean Label Ingredient Market—Growth, Trends, and Forecast (2018–2023)". [(accessed on 12 October 2021)]. Available online: https://www.researchandmarkets.com/research/wp8cd3/global_clean?w=12
- 194. Shaltout, F. A.; Elshater, M. and Wafaa, Abdelaziz (2015): Bacteriological assessment of street vended meat products sandwiches in Kalyobia Governorate. Benha Vet. Med.J.28 (2):58-66.
- 195. Luong N.-D.M., Coroller L., Zagorec M., Membré J.-M., Guillou S. Spoilage of chilled fresh meat products during storage: A quantitative analysis of literature data. Microorganisms. 2020;8:1198.
- 196. Shaltout, F. A.; Gerges, M.T. and Shewail, A.A.(2018):Impact of Organic Acids and Their Salts on Microbial Quality and Shelf Life of Beef. Assiut veterinary medical journal 64(159): 164-177
- 197. Martillanes S., Rocha-Pimienta J., Llera-Oyola J., Gil M.V., Ayuso-Yuste M.C., García-Parra J., Delgado-Adámez J. Control of Listeria monocytogenes in sliced dry-cured Iberian ham by high pressure processing in combination with an ecofriendly packaging based on chitosan, nisin and phytochemicals from rice bran. Food Control. 2021;124:107933.
- 198. Shaltout,F.A.;Ghoneim, A.M.; Essmail, M.E. and Yousseif,A.(2001): Studies on aflatoxin B1 residues in rabbits and their pathological effects. J.Egypt. Vet. Med. Association 61(2):85-103.
- 199. Lee J.-S., Choi Y.S., Lee H.G. Synergistic antimicrobial properties of nanoencapsulated clove oil and thymol against oral bacteria. Food Sci. Biotechnol. 2020;29:1597–1604.
- 200. Shaltout,F.A. and Hanan ,M.T. El-Lawendy (2003): Heavy Metal Residues In Shawerma. Beni-Suef Vet.Med.J. 13(1):213-224.
- 201. Park S., Mun S., Kim Y.-R. Influences of added surfactants on the water solubility and antibacterial activity of rosemary extract. Food Sci. Biotechnol. 2020;29:1373–1380.

202. Shaltout, F.A. and Hashim, M.F. (2002): Histamine in salted, Smoked and Canned Fish products. Benha Vet. Med.J.13 (1):1-11.

- 203. Fang Z., Zhao Y., Warner R.D., Johnson S.K. Active and intelligent packaging in meat industry. Trends Food Sci. Technol. 2017;61:60–71.
- 204. Shaltout, F.A.; Hashim, M.F. and Elnahas, s. (2015): Levels of some heavy metals in fish (tilapia nilotica and Claris lazera) at Menufia Governorate. Benha Vet. Med. J. 29 (1):56-64.
- 205. Yong H.I., Kim T.K., Choi H.D., Jang H.W., Jung S., Choi Y.S. Clean label meat technology: Pre-converted nitrite as a natural curing. Food Sci. Anim. Resour. 2021;41:173–184.
- 206. Shaltout,F.A. and Ibrahim, H.M.(1997): Quality evaluation of luncheon and Alexandrian sausage. Benha Vet. Med.J.10 (1):1-10.
- 207. Barcenilla C., Ducic M., López M., Prieto M., Álvarez-Ordóñez A. Application of lactic acid bacteria for the biopreservation of meat products: A systematic review. Meat Sci. 2021;183:108661.
- 208. Shaltout, F.A.; Nassif, M and Shakran, A(2014): Quality of battered and breaded chicken meat products. Global Journal of Agriculture and Food Safety Science 1(2) ISSN 2356-7775.
- 209. Crowe W., Elliott C.T., Green B.D. A review of the in vivo evidence investigating the role of nitrite exposure from processed meat consumption in the development of colorectal cancer. Nutrients. 2019;11:2673.
- 210. Shaltout, F.A., Amani M. Salem, A. H. Mahmoud, K. A(2013): Bacterial aspect of cooked meat and offal at street vendors level .Benha veterinary medical journal, 24(1): 320-328.
- 211. Choe E. Roles and action mechanisms of herbs added to the emulsion on its lipid oxidation. Food Sci. Biotechnol. 2020;29:1165–1179.
- 212. Shaltout, F.A. and Salem, R.M. (2000): Moulds, aflatoxin B1 and Ochratoxin A in Frozen Livers and meat products. Vet . Med. J. Giza 48(3):341-346.
- 213. Marrone R., Smaldone G., Ambrosio R.L., Festa R., Ceruso M., Chianese A., Anastasio A. Effect of beetroot (Beta vulgaris) extract on black angus burgers shelf life. Ital. J. Food Saf. 2021;10:9031.
- 214. Yasser H. Al-Tarazi, A. Al-Zamil, Shaltout FA. and H. Abdel-Samei (2002). Microbiological status of raw cow milk marketed in northern Jordan. AVMJ Volume 49 Issue 96 Pages 180-194
- 215. Lee N.-K., Paik H.-D. Status, Antimicrobial mechanism, and regulation of natural preservatives in livestock food systems. Korean J. Food Sci. Anim. Resour. 2016;36:547–557.
- 216. Shaltout FA, Zakaria IM and Nabil ME.(2018): Incidence of Some Anaerobic Bacteria Isolated from Chicken Meat Products with Special Reference to Clostridium perfringens. Nutrition and Food Toxicology2(5):429-438.
- 217. Olszewska M.A., Gędas A., Simões M. Antimicrobial polyphenol-rich extracts: Applications and limitations in the food industry. Food Res. Int. 2020;134:109214.
- 218. Shaltout, F. A.; El-diasty, E.M. and Mohamed, M. S.(2014): Incidence of lipolytic and proteolytic fungi in some chicken meat products and their public health significance. 1st Scientific conference of food safety and Technology .2014, pp. 79-89.
- 219. Chaleshtori F.S., Arian A., Chaleshtori R.S. Assessment of sodium benzoate and potassium sorbate preservatives in some products in Kashan, Iran with estimation of human health risk. Food Chem. Toxicol. 2018;120:634–638.
- 220. Shaltout, F. A.; El-diasty, E.M.; Salem, R. M. and Asmaa, M. A. Hassan. 2016: Mycological quality of chicken carcasses and extending shelf-life by using preservatives at refrigerated storage. Veterinary Medical Journal Giza, 62(3):1-10.
- 221. International Agency for Research on Cancer (IARC) monographs on the evaluation of carcinogenic risks to humans Ingested nitrate and nitrite, and cyanobacterial peptide toxins. IARC Monogr. Eval. Carcinog. Risks Hum. 2010;94:1–412.

222. Shaltout FA, R.M. Salem, E.M. El-Diasty and W.I.M. Hassan. 2019: Effect of Lemon Fruits and Turmeric Extracts on Fungal Pathogens in Refrigerated Chicken Fillet Meat. Global Veterinaria 21 (3): 156-160,

Page 14 of 15

- 223. Cao Q., Yan J., Sun Z., Gong L., Wu H., Tan S., Lei Y., Jiang B., Wang Y. Simultaneous optimization of ultrasound-assisted extraction for total flavonoid content and antioxidant activity of the tender stem of Triarrhena lutarioriparia using response surface methodology. Food Sci. Biotechnol. 2021;30:37–45.
- 224. Shaltout FA, El-diasty, E,M. ;Elmesalamy, M. and Elshaer, M.(2014): Study on fungal contamination of some chicken meat products with special reference to 2 the use of PCR for its identification. Conference, Veterinary Medical Journal Giza vol. December 2014/12/17 vol.60 1-10.
- 225. Piper J.D., Piper P.W. Benzoate and sorbate salts: A systematic review of the potential hazards of these invaluable preservatives and the expanding spectrum of clinical uses for sodium benzoate. Compr. Rev. Food Sci. Food Saf. 2017;16:868–880.
- 226. Shaltout, F. A.; Salem, R. M; El-diasty, Eman and Fatema, A.H. Diab. (2016): Mycological evaluation of some ready to eat meat products with special reference to molecular characterization. Veterinary Medical Journal Giza. 62(3): 9-14.
- 227. Shim S.-M., Seo S.H., Lee Y., Moon G.-I., Kim M.-S., Park J.-H. Consumers' knowledge and safety perceptions of food additives: Evaluation on the effectiveness of transmitting information on preservatives. Food Control. 2011;22:1054–1060.
- 228. Shaltout FA, Ahmed, A.A. Maarouf, Eman, M.K. Ahmed(2018): Heavy Metal Residues in chicken cuts up and processed chicken meat products. BENHA VETERINARY MEDICAL JOURNAL, 34(1): 473-483.
- 229. Ministry of Food and Drug Safety (MFDS) Food Additives Code. [(accessed on 12 October 2021)]. Available online: https://www.mfds.go.kr/eng/brd/m 15/view.do?seq=72432
- 230. Shaltout ,F.A.; Hanan M. Lamada , Ehsan A.M. Edris.(2020): Bacteriological examination of some ready to eat meat and chicken meals. Biomed J Sci & Tech Res., 27(1): 20461-20465.
- 231. Matthews K.R., Kniel K.E., Montville T.J. Food Microbiology: An Introduction. 4th ed. ASM Press; Washington, DC, USA: 2017.
- 232. Sobhy, Asmaa and Shaltout, Fahim(2020): Prevalence of some food poisoning bacteria in semi cooked chicken meat products at Qaliubiya governorate by recent Vitek 2 compact and PCR techniques. Benha Veterinary Medical Journal 38 (2020) 88-92.
- 233. Bohrer B.M. Nutrient density and nutritional value of meat products and non-meat foods high in protein. Trends Food Sci. Technol. 2017;65:103–112.
- 234. Sobhy, Asmaa and Shaltout, Fahim(2020): Detection of food poisoning bacteria in some semi-cooked chicken meat products marketed at Qaliubiya governorate. Benha Veterinary Medical Journal 38 (2020) 93-96.
- 235. Zhou G., Xu X., Liu Y. Preservation technologies for fresh meat–A review. Meat Sci. 2010;86:119–128.
- 236. Shaltout, F.A.(2024): Abattoir And Bovine Tuberculosis as A Reemerging Foodborne Diseas. Clinical Medical Reviews and Report 6(1):1-7.
- 237. World Health Organization . WHO Estimates of the Global Burden of Foodborne Diseases: Foodborne Disease Burden Epidemiology Reference Group 2007–2015. World Health Organization (WHO); Geneva, Switzerland: 2015. pp. 1–15.
- 238. Shaltout, F.A.(2023): Viruses in Beef, Mutton, Chevon, Venison, Fish and Poultry Meat Products. Food Science & Nutrition Technology 8(4):1-10.
- 239. Lee H., Yoon Y. Etiological agents implicated in foodborne illness worldwide. Food Sci. Anim. Resour. 2021;41:1–7.

240. Farnaud S., Evans R.W. Lactoferrin—A multifunctional protein with antimicrobial properties. Mol. Immunol. 2003;40:395–405.

Ready to submit your research? Choose ClinicSearch and benefit from:

- > fast, convenient online submission
- > rigorous peer review by experienced research in your field
- > rapid publication on acceptance
- > authors retain copyrights
- > unique DOI for all articles
- > immediate, unrestricted online access

At ClinicSearch, research is always in progress.

Learn more https://clinicsearchonline.org/journals/clinical-oncology-case-reports



© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.