

Bioactive profiling of essential oil of Terminalia arjuna stem bark collected from Orathur village, Tamilnadu, India

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Abstract

Bioactive profiling of Terminalia arjuna stem bark essential oil by GC/MS intends to showcase the medicinal properties and characterization of bioactive compounds. Bioactive profiling of essential oils from Terminalia arjuna stem bark revealed the presence of 31 bioactive compounds with their retention time. Cyclohexylhexanoate (10.78 %), D-limonene (9.57 %), ethyltrans-4-decenoate (9.52 %), α -himachalene (7.21 %), β -sesquiphellandrene (6.09 %), β -caryophyllene (5.66 %), Trans-2-Tetradecen-1-ol (4.09 %), β -Guaiene (4.02 %), 2-methyldecahydronaphthalene (3.72 %), cis-7-hexadecane (3.11 %), α -cadinol (3.04 %), 1-octanal (2.57 %) and ethylbenzene (2.02 %) were the major compounds above 2 % while compounds less than 2.0 % includes, 3-Hexenylhexanoate (0.97 %), 2,6,11-Trimethyldodecane (1.36 %), 2,3,6,7-Tetramethyloctane (0.25 %), β -Selinenol (1.77 %), (-) δ -Cadinol (0.01 %), Cubenol (0.03 %), α -Bisabolol (0.04 %), α -Himachalene (1.88 %), 1,3,5,8-Undecatetraene (1.02 %), Ethyltrans-4-Decenoate (0.05 %), α -Terpinolene (0.94 %), Trans-2-Nonenal (0.06 %), Geranyl Acetone (1.67 %), Cis-6-Pentadecen-1-ol (0.51 %) and Hexahydrofarnesol (0.87 %). It was concluded that essential oil from Terminalia arjuna stem bark is rich in several phytochemicals with medicinal properties and can be used to reduce the increasing cases of antimicrobial resistance.

Keywords: terminalia arjuna; phytochemicals; safety; medicine; antimicrobial; resistance

Introduction

Terminalia arjuna is an evergreen plant belonging to the family Combretaceae (Kapoor et al., 2014). The tree is found in most parts of India, Sri Lanka, China, Pakistan, Bangladesh and Malaysia (Pashazanousi et al., 2012). The tree can grow up to 30 meters in height and is highly medicinal due to the presence of tannins, alkaloids, flavonoids, saponins, glycosides, phenolic compounds amongst others (Saha et al., 2012; Bharani et al., 2004). These phyto-components performs numerous biological functions including, anti-inflammatory (Alagbe et al., 2021), antifungal, antiviral, antimicrobial, immune stimulator, cytotoxic, gastro-protective, anti-ulcer, anti-diabetic, hypolipidemic, antioxidant, osteogenic, anti-helminthic and cardio-protective amongst others (Bharani et al., 2004; Paul et al., 2016). The plant parts (leaves, stem bark and root extracts) has reportedly been used for the treatment of severe diarrhoea and dysentery, urethral discharge, gastrointestinal infection, chest, pain, waist pain, irregular menstruation, internal pile, malarial, quick ejaculation, headache, hypertension, dysentery, premature aging, memory improvement, blood cleansing, chronic venous, insufficiency, mental function, minor burns, scars, skin ulcers, varicose veins, wound healing, rheumatism and congestive heart failure (Khalil, 2005; Bharani, et al., 1995).

The stem bark from the plant is characterized with sweet, cooling, styptic, tonic, anti-dysenteric, and febrifuge properties (Desai et al., 2015). Extracts from the leaves and roots of Terminalia arjuna can inhibit the growth of some pathogenic organisms such as, Pseudomonas aeruginosa, Staphylococcus aureus, Escherichia coli, Bacillus subtilis and Candida albicans (Yaidikar and Thakur, 2015). According to Singh et al. (2022; Alagbe, 2023), concentration of phytochemicals in medicinal plants can be influenced by age of plant, specie, geographical location, extraction technique amongst others. These phyto-components have been reported to be safe, environmentally friendly and has no withdrawal period (Alagbe et al., 2020). However, errors in botanical identification, Interference of medicinal plants and conventional pharmacological therapy and dearth of reports on the side effects of medicinal plants can cause toxicity in phytomedicine in human and animals (Olujimi et al., 2024).

Therefore, this study was carried out to determine the bioactive profiling of essential oil of Terminalia arjuna stem bark collected from Orathur village, Tamilnadu, India

Materials and methods

Description of experimental area

The experiment was carried out at the department of Biochemistry, Sumitra Research Institute, Gujarat located between 28° 20' N and 75° 30' East India in the months of August to October, 2022.

Collection and extraction of essential oil from *Terminalia arjuna* stem bark

Fresh mature stem bark from *Terminalia arjuna* was collected from different sites at Orathur village, Tamilnadu, India and sent to taxonomy department of the same institute for proper authentication before it was assigned an

identification number (HF/008C/2023). Extraction of essential oil from *Terminalia arjuna* stem bark was carried out using steam distillation technique with Clevenger apparatus. Extracted oil was sent to the laboratory for further analysis.

Bioactive profiling of essential oil from *Terminalia arjuna* stem bark

Bioactive profiling of essential oil from *Terminalia arjuna* stem bark was carried out using Lauret gas chromatography - mass spectrometry (Model FG/008, Netherlands). Identification of each bioactive compound was carried out by comparing their mass spectra with those of reference compounds from the Library of National Institute of Standard and Technology (NIST, 2011) database.

S/N	Compounds	Reaction time (min)	% Area
1	3-Hexenylhexanoate	5.62	0.97
2	2,6,11-Trimethyldodecane	6.27	1.36
3	β -Caryophyllene	6.33	5.66
4	Cyclohexylhexanoate	7.07	10.78
5	γ -Cadinene	7.55	2.67
6	β -Sesquiphellandrene	7.92	6.09
7	β -Linalool	8.09	2.51
8	D-Limonene	8.47	9.57
9	2,3,6,7-Tetramethyloctane	8.84	0.25
10	β -Selinol	8.93	1.77
11	α -Cadinol	9.62	3.04
12	(-) δ -Cadinol	9.95	0.01
13	α -Bisabolol	10.50	0.04
14	Cubenol	11.10	0.03
15	α -Himachalene	11.55	7.21
16	β -Guaiene	12.35	4.02
17	α -Himachalene	12.67	1.88
18	1,3,5,8-Undecatetraene	12.85	1.02
19	Ethyltrans-4-Decenoate	13.06	0.05
20	α -Terpinolene	14.54	0.94
21	1-Octanal	15.12	2.57
22	1,8-Cineole	15.76	3.5
23	Ethyltrans-4-Decenoate	16.27	9.52
24	2-methyldecahydronaphthalene	17.16	3.72
25	Ethylbenzene	18.09	2.02
26	Trans-2-Nonenal	19.22	0.06
27	Geranyl Acetone	19.85	1.67
28	Cis-6-Pentadecen-1-ol	20.06	0.51
29	Trans-2-Tetradecen-1-ol	21.38	4.09
30	Cis-7-Hexadecane	22.40	3.11
31	Hexahydrofarnesol	22.75	0.87
	Total	91.51	
	Number of compounds		
	Monoterpenes	27.51	
	Diterpenes	7.96	
	Triterpenes	1.03	
	Sesquiterpenes	-	
	Non-terpenes	55.01	

Table 1: Bioactive profiling of *Terminalia arjuna* stem bark essential oil by GC/MS

Results And Discussion

Bioactive profiling of essential oils from *Terminalia arjuna* stem bark revealed the presence of 31 bioactive compounds with their retention time. Cyclohexylhexanoate (10.78 %), D-limonene (9.57 %), ethyltrans-4-decenoate (9.52 %), α -himachalene (7.21 %), β -sesquiphellandrene (6.09 %), β -caryophyllene (5.66 %), Trans-2-Tetradecen-1-ol (4.09 %), β -Guaiene (4.02 %), 2-methyldecahydronaphthalene (3.72 %), cis-7-hexadecane (3.11 %), α -cadinol (3.04 %), 1-octanal (2.57 %) and ethylbenzene (2.02 %) were the major compounds above 2 % while other compounds reported were less than 2.0 %. It is worthy to note that all these bioactive compounds or phytochemicals possess medicinal or therapeutic properties (Alagbe and

Shittu, 2020; Alagbe et al., 2021). This result aligns with a previous study by Kokkiripati et al. (2013); Hafiz et al. (2014); Chaudhari and Mengi (2006). For instance, cyclohexylhexanoate, β -caryophyllene, β -Linalool, β -sesquiphellandrene, β -selinenol and α -cadinol have been reported to possess antimicrobial, antifungal, antidiarrhoea, antibacterial, anticancer, antioxidant and anti-helminthic properties (Subavathy and Thilaga, 2015; Mangrove et al., 2014). 2, 6, 11-trimethyldodecane, 2, 3, 6, 7-tetramethyloctane, α -himachalene, α -terpinolene and cis-6-pentadecen-1-ol have antimicrobial and gastro-protective activities (Doughari, 2012; Olajuyige et al., 2011). α -bisabolol, 2-methyldecahydronaphthalene, ethyltrans-4-decenoate have been suggested to have antibacterial and cardio-protective effects (Devendran and Ba;asubramanian, 2011; Lima et al., 2010). Trans-2-nonenal, geranyl acetone and ethylbenzene have antifungal and antidiarrhoea properties (Mamza et al., 2012; Awa et al., 2012). Screening of bioactive compounds from herbal plants can lead to the discovery of new medicinal drugs which have efficient protection and treatment roles against various diseases (Soma et al., 2010; Alagbe et al., 2024). The concentrations of phytochemicals in herbal plants can be influenced age of plant, geographical location, specie, processing methods amongst others (Alagbe et al., 2023a; Alagbe et al., 2023b).

Conclusion

Naturally, medicinal plants are loaded with phytochemicals with medicinal properties. These compounds can perform numerous biological roles such as, antimicrobial, antifungal, anti-helminthic, hepato-protective, immunostimulatory, cytotoxic, antioxidant, antiviral amongst others.

References

1. Alagbe, J. O (2024). Proximate, mineral and phytochemical analysis of some medicinal plants collected from Orathur village, Thiruporur Taluk Kancheepuram district Tamilnadu, India. *World Journal of Agriculture and Forestry Sciences*, 2(2): 30-35.
2. Mamza, U.T., Sodipo, O.A. & Khan, I.Z. (2012). Gas Chromatography - Mass spectrometry (GC - MS) analysis of bioactive components of *Phyllanthus amarus* Leaves. *International Research Journal of Plant Science*, 3 (10), 208 - 215
3. Awa, E.P., Ibrahim, S. & Ameh, D.A. (2012). GC/MS Analysis and Antimicrobial Activity of Dimethyl Ether fraction of Methanolic Extract from stem Bark of *Annona senegalensis* Pers. *International Journal of Pharmaceutical Sciences and Research*. 3 (11), 4213 – 4218
4. Devendran, G. & Balasubramanian, U. (2011). Qualitative phytochemical screening and GC-MS analysis of *Ocimum Santum* L. leaves. *Asian Journal of Plant Science and Research*, 1 (4), 44- 48.
5. Lima, A.L., Parial, R., Das, M. & Das, A.K. (2010). Phytochemical and Pharmacological studies of ethanolic extract from the leaf of mangrove plant *Phoenix paludosa* Roxb. *Malaysian Journal of Pharmaceutical Sciences*, 8 (2), 59-69.
6. Doughari, J.H. (2012). *Phytochemicals: Extraction Methods, Basic Structures and Mode of Action as Potential Chemotherapeutic Agents- A Global Perspective of Their Role in Nutrition and Health*. Dr. Venketeshwer Rao. Ed). ISBN: 978-953-51-0296-0.

7. Olajuyige, O.O., Babalola, A.E. & Afolalayan, A.J. (2011). Antibacterial and phytochemical screening of crude ethanolic extracts of *Waltheria* Linn. *African Journal of Microbiology Research*, 5 (22), 3760-3764.
8. Subavathy, P. & Thilaga, R.D. (2015). GC-MS Analysis of Bioactive Compounds from Whole Body of Methanolic Extract of *Cypraea arabica*. *World Journal of Pharmaceutical Research*, 5 (3), 800-806.
9. Mangrove-Abayomi, O.E., Kenneth, E and Mkaparu, K.I. (2014). Chemometric profiling of methanolic leaf extract of *Cinddoscolus aconitifolius* (Euphorbiaceae) using UV-VIS, FTIR and GC-MS techniques. *Peak journal of Medicinal Plant Research*, 2 (1), 6-12.
10. Alagbe, J.O., Bamigboye, S., Nwosu, G.C., Agbonika, D.A and Kadiri Mercy Cincinsoko. (2023). Characterization of bioactive compounds in *Luffa aegyptiaca* leaf ethanolic extracts using gas chromatography and mass spectrometry (GC-MS). *Drug Discovery*; 17:101011.
11. Alagbe, J.O., Kadiri, M.C., Oluwafemi, R.A., Agubosi, O.C.P and Anorue, D.N. (2023b). Analysis of bioactive compounds in ethanolic extracts of *Xylopi aethiopica* leaves using gas chromatography and mass spectrometry technique. *American Journal of Science on Integration and Human Development*, 1(1): 1-10.
12. Alagbe, J.O. (2023). Bioactive compounds in ethanolic extract of *Strychnos innocua* root using gas chromatography and mass spectrometry (GC-MS). *Drug Discovery*, 17:41005.
13. Singh Sharma., Alagbe Olujimi John., Liu Xing., Sharma Ram and Kumar Amita (2022). Comparative analysis of ethanolic *Juniperus thurifera* leaf, stem bark and root extract using gas chromatography and mass spectroemetry. *International Journal of Agriculture and Animal Production*, 2(6): 18-27.
14. Alagbe, J.O., Adedeji, M.O., Habiba, Z., Nwosu, Gloria and Wyedia Dabara Comfort (2021). Physico-chemical properties of *Indigofera zollingeriana* seed oil. *Asian Journal of Advances in Medical Science* 3(4): 306-308.
15. Alagbe, J.O., Shittu, M.D and Ushie, F.T. (2021). GC-MS analysis of methanolic stem bark extract of *Zollingeriana indigofera*. *Asian Journal of Advances in Research* 11(4): 144-146.
16. Alagbe, J.O (2020). Chemical evaluation of proximate, vitamin and amino acid profile of leaf, stem bark and roots of *Indigofera tinctoria*. *International Journal on Integrated Education*. 3(10): 150-157.
17. Shittu, M.D and Alagbe, J.O. (2020). Phyto-nutritional profiles of broom weed (*Sida acuta*) leaf extract. *International Journal of Integrated Education*. 3(11): 119-124
18. Kapoor D, Vijayvergiya R, Dhawan V. (2014). *Terminalia arjuna* in coronary artery disease: Ethnopharmacology, pre-clinical, clinical and safety evaluation. *Journal of Ethnopharmacology*, 155:1029-1045.
19. Pashazanousi MB, Raeesi M, Shirali S. (2012). Chemical composition of the essential oil, antibacterial and antioxidant activities, total phenolic and flavonoid evaluation of various extracts from leaves and fruit peels of *Citrus limon*. *Asian Journal of Chemistry*, 24:4331-4.
20. Hafiz FB, Towfique NM, Sen MK, Sima SN, Azhar BS, Rahman MM. (2014). A comprehensive ethno-pharmacological and phytochemical update review on medicinal plant of *Terminalia arjuna* Roxb. of bangladesh. *Sch Acad J Pharmacology*, 3:19-25
21. Chaudhari M, Mengi S. (2006). Evaluation of phytoconstituents of *Terminalia arjuna* for wound healing activity in rats. *Phytother Research*, 20:799-805.
22. Kokkiripati PK, Kamsala RV, Bashyam L, Manthapuram N, Bitla P, Peddada V. (2013). Stem-bark of *Terminalia arjuna* attenuates human monocytic (THP-1) and aortic endothelial cell activation. *J Ethnopharmacology*, 146:456-464

23. Olujimi, J.O., Anuore, D.N and Aliyu, K.I. (2024). Chrysophyllum albidum stem bark powder: effects on performance and carcass characteristics of Japanese quails. *World Journal of Clinical Studies*, 2(2): 41-48.
24. Saha A, Pawar VM, Jayaraman S. (2012). Characterization of polyphenols in Terminalia arjuna bark extract. *Indian J Pharm Science*, 74:339-347
25. Bharani A, Ahirwal K, Jain N. (2004). Terminalia arjuna reverses impaired endothelial function in chronic smokers. *Indian Heart Journal*, 56:123-128.
26. Paul S, Ghosh D, Ghosh AK, Bhowmick D, Bandyopadhyay D, Chattopadhyay A. (2016). Aqueous bark extract of Terminalia arjuna protects against phenylhydrazine induced oxidative damage in goat red blood cell membrane bound and metabolic enzymes. *Int J Pharm Pharm Science*, 8:62-70.
27. Khalil S. (2005). Effect of Statin Versus Terminalia arjuna on Acute Myocardial Infarction (DNB thesis). New Delhi, India: National Board of Examination.
28. Desai SD, Patil BS, Kanthe PS, Potekar RM. (2015). Effect of ethanolic extract of Terminalia arjuna on liver functions and histopathology of liver in albino rats fed with hyperlipidemic diet. *Int J Pharm Pharm Science*, 7:302-306.
29. Bharani A, Ganguli A, Bhargava KD. (1995). Salutary effect of Terminalia arjuna in patients with severe refractory heart failure. *Int J Cardiology*, 49:191-199.
30. Yaidikar L and Thakur S. (2015). Arjunolic acid, a pentacyclic triterpenoidal saponin of Terminalia arjuna bark protects neurons from oxidative stress associated damage in focal cerebral ischemia and reperfusion. *Pharmacol Repository*, 67:890-895

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