

The Impact of Bisphenol A on Gonadal Hormones and Histological Structure of Wistar rats

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Abstract:

Introduction: Bisphenol A (BPA), a chemical found in plastics and consumer goods, significantly impacts fertility in female rats, disrupting oocyte maturation, ovulation, and sperm quality, leading to impaired implantation and embryonic development. Further research is needed to develop strategies to mitigate BPA's reproductive toxicity.

Materials and methods: The study involved 40 Wistar rats from the University of Khartoum, Sudan, who were given Extra Pure Bisphenol A powder for four weeks. After anesthesia and slaughter, their blood was analyzed for sexual hormones and lesions.

Results: The study investigated the impact of BPA on rats' reproductive hormones, showing significant changes in both control and treated groups, including reduced FSH, LH, testosterone, estrogen, progesterone, and Prolactin levels, and abnormal ovaries.

Conclusion: BPA exposure reduced reproductive hormones in rats, with elevated Prolactin levels. Males experienced seminiferous epithelium degeneration, sperm reduction, and abnormal antral, cystic, and atretic follicles.

Keywords: bisphenol a; estrogen; progesterone; prolactin; reproductive system; testosterone; wistar rats

1. Introduction

Bisphenol A (BPA), recognized as an endocrine-disrupting chemical, is notably prevalent in the environment and has found extensive application in the manufacturing of plastics, epoxy resins, and various consumer goods [1]. Owing to its omnipresent nature and potential repercussions on health, apprehensions about BPA exposure and its implications for the well-being of both humans and animals have been on the rise. A multitude of research endeavors have delved into the possible reproductive hazards associated with BPA, particularly emphasizing its effects on fertility [2]. One critical area impacted by BPA exposure is oocyte maturation and ovulation in female rats [3]. Research has shown that BPA can disturb the maturation of oocytes and the process of ovulation, resulting in a significant decrease in ovulation rates and ultimately reducing fertility levels among the exposed subjects. Similarly, in male rats, BPA has been linked to a decline in sperm quality and function [4]. Specifically, exposure to BPA is associated with impaired sperm quality, reduced sperm motility, and a decrease in sperm count [5-6].

Moreover, the effects of BPA extend to implantation and embryonic development. Studies have indicated that BPA exposure can hinder successful implantation and proper embryonic development, potentially leading to increased instances of miscarriage and the occurrence of fetal abnormalities in affected individuals [7]. In addition to these direct impacts on fertility, BPA-induced hormonal deregulation further exacerbates reproductive challenges. By disrupting the production and regulation of crucial reproductive hormones such as estrogen, testosterone, and follicle-stimulating hormone (FSH), BPA perpetuates the adverse effects on overall reproductive function -- underscoring the urgency of understanding and mitigating the consequences of BPA exposure on reproductive health [8].

The current research aimed to study the impact of exposure to small doses of BPA to Wistar rats of both sexes on the gonadal hormones and histological structure

2. Materials and methods

The present study was carried out in the Department of Biochemistry and Molecular Biology, Faculty of Science and Technology, El Neelain, University, Sudan, after getting approval from Scientific Research Ethical Committee. Forty Wistar rats were obtained from the Faculty of Pharmacy University of Khartoum, reared within the premises of the animal house under 12 hours' photoperiod with standard feed and drinking water provided *ad libitum* before the commencement of experimental feeding. Room temperature was maintained at 25 ± 2 °C at adequate house ventilation.



Figure 1: Wister rats

2.1 Data collection

2.1.1 Serum analysis

After the end of the experimental period, rats of the control and treatment groups were anaesthetized with diethyl ether and humanely slaughtered. Blood was collected at slaughter in clean sterile vials and sera were separated thereafter to be analyzed for the sexual hormones, Follicle-Stimulating Hormone (FSH), Leutinizing Hormone (LH), Testosterone, Progesterone, Estrogen and Prolactin according to Aviva Systems Biology [9].

2.1.2 Histopathological methods

Necropsy was conducted to identify gross lesions and specimens of tests and ovaries were immediately being collected immediately after slaughter of rats, fixed in 10% neutral buffered formalin and embedded in paraffin wax, sectioned at 5m and stained with Haematoxylin and Eosin (H & E) [10].

2.2. Statistical analysis

Mean values of Testosterone, Prostaglandin, Progesterone, Estrogen and Prolactin concentration, were compared using student's t-test [11].

3. Results:

3.1. Clinical observations

The control Group 1 remained clinically normal throughout the experimental period. On the fourth day of the experiment, rats of group 2 showed nervous signs (aggressiveness and defensive behavior) and dosing resistance.

Then the animals were randomly allotted into four groups 1, 2, 3, and 4 each of ten rats. Group 1 was designated as the male control group, group 2 was designed as the male bisphenol-treated group. Group 3 is the female control group, whereas Group 4 is the female bisphenol A- treated group. Extra Pure (97%) Bisphenol A powder (Sangon, China) was thoroughly dissolved in distilled water and rats of group 2 (male rats) and 4 (female rats) received this test chemical by oral gavage dose at $25 \mu\text{g}/\text{kg}$ body weight/day for four weeks period.

3.2. Serum sex hormones concentration

Changes in concentrations of serum reproductive hormones are presented in Table 1. In male and female groups exposed to $25 \mu\text{l}/\text{kg}/\text{day}$ BPA, all the measured hormones were altered. Serum concentrations of FSH were reduced by 36.9% and 69.5% in BPA treated male and female, respectively and those of LH were reduced by 48.1% and 43.4%, respectively. Serum concentrations of testosterone were significantly reduced ($P < 0.05-0.01$) in the two treatment groups. No estrogen was detected in both male control and treated groups, while the concentration in the female rats was significantly decreased ($P < 0.05$) in BPA treated group. Significant decrease ($P < 0.05$) in progesterone concentration was observed in female treated rats, but that of the male treatment group did not change. Significantly ($P < 0.05-0.01$) higher concentration of Prolactin was recorded in both male and female treated rats than the controls.

3.3. Histopathological findings

Testis and ovaries of the male and female control groups remain normal throughout the experimental period. Significant histopathological alterations were observed in the treated group. Marked reduction of the thickness of the seminiferous tubules, along with an increased degeneration of the seminiferous epithelium was visualized. Associated with these alterations, a significant reduction in the number of sperms in the lumen of the seminiferous tubule was found (Fig. 2). Ovaries of treated female rats given BPA at $25 \mu\text{g}/\text{kg}$ body weight/day showed lack of normal appearing antral follicles, enlarge cystic follicles and showing a number of atretic follicles (Fig. 3).

Rat Sex	BPA Dose 25 $\mu\text{g}/\text{kg}/\text{day}$	FSH mIU/mL	LH mIU/mL	Testosterone ng/mL	Estrogen pg/mL	Progesterone ng/mL	Prolactin ng/mL
Male	Control M	5.85 ± 0.16	4.10 ± 0.05	7.06 ± 0.08	ND	1.52 ± 0.23	4.17 ± 0.07
	Treated M	$2.16 \pm 0.02^*$	$1.97 \pm 0.16^*$	$2.13 \pm 0.04^{**}$	ND	$2.15 \pm 0.10^{\text{NS}}$	$7.26 \pm 0.12^*$
Female	Control F	8.24 ± 0.16	7.49 ± 0.05	1.64 ± 0.12	10.51 ± 0.57	5.61 ± 0.40	2.30 ± 0.12

	Treated F	5.73±0.13*	3.25±0.02**	0.65±0.14 *	7.10±0.12*	2.4±0.15*	8.54±0.29**
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Values are means ±SE, NS = not significant, *Denotes mean values significant at (P<0.05), **Significant= (P<0.01),

M= Male, F= Female

Table 1: Effects of BPA at 25 µg/kg/day on male and female reproductive hormones

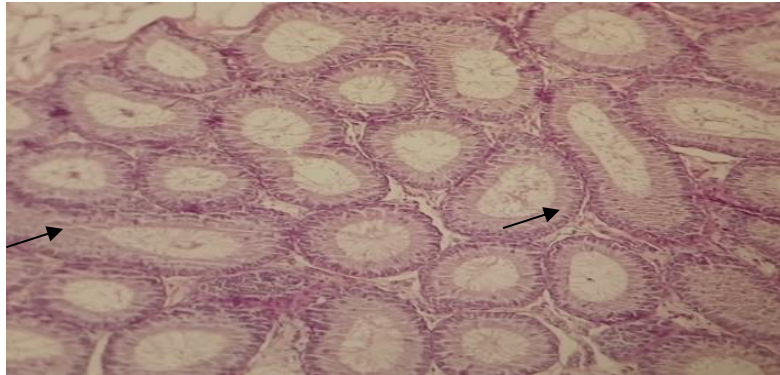


Figure 2: Reduction of the thickness and an increase in the degeneration of the seminiferous epithelium as well as a reduction in the number of sperms in the lumen of the seminiferous tubule. H&E x100

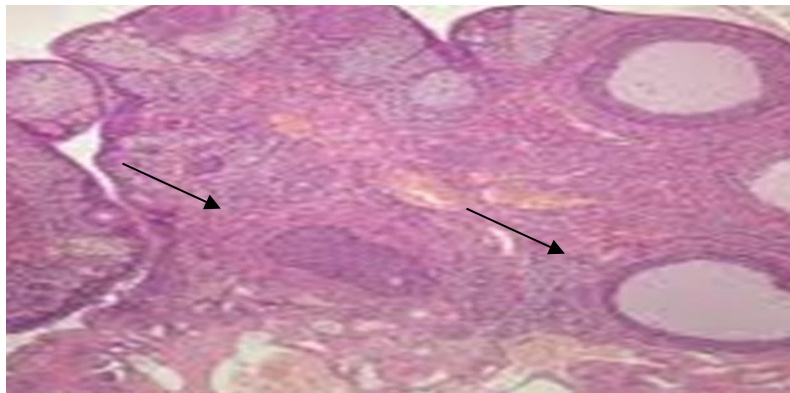


Figure 3: Lack of normal appearing antral follicles, enlarge cystic follicles and showing a number of atretic follicles in a female rat receiving oral BPA at 25 µg/kg H&E x100.

4. Discussion

The results of the present study have indicated that the serum hormonal levels of FSH, LH, Testosterone, Estrogen and Progesterone of female Wistar rats were significantly decreased with a significant increase in prolactin in the test group when BPA was orally administered at a dose of 25 µg/kg/day. A study by [12] revealed that non-pregnant female Wistar rats exposed to different oral doses of BPA (5, 50, 300, 600 and 800 mg/kg body weight /week) exhibited significant decreased levels of LH, FSH, E2, PROG and PRL with altering the mating activity, reducing the reproductive capacity and leading to decrease female fertility and weight of reproductive organs (ovary and uterus). [13] Study findings indicated that Adult female Sprague Dawley rats orally exposed to 330 mg/kg BW of BPA for 10 and 12 weeks revealed reduction in estrogen and progesterone concentration [14] found that adult female rats exposed to 0.1 mg/kg BW of BPA for 90 days showed decreased estradiol concentration. This result might be attributed to the effect of BPA that causes down regulation of P450 aromatase mRNA expression in granulosa cells [15] that play an important role in estrogen biosynthesis [16] or it might be due to the reduction in steroidogenic acute regulatory protein [17], which is a critical steroidogenic protein responsible for the transfer of cholesterol from the outer to the inner mitochondrial membrane during androgen biosynthesis [18]. On the other hand, bisphenol A disrupts estrogen hormone, and exposure to BPA resulted in increased atresia of the ovarian follicles [19] Delclos *et al.* [20] observed

no significant treatment-related effects on serum hormone levels in the low BPA dose range (2.5–2700 µg/kg/day) in females, but serum estradiol was significantly increased by 67% and 113%, respectively in the 100,000 and 300,000 µgBPA/kg/day exposures. Progesterone was significantly decreased by 51% in the highest BPA dose (300,000 µg BPA/kg /day). The highest dose of BPA had significantly elevated prolactin levels, although they did not differ significantly from the control females. No significant differences in FSH and LH levels were observed in 100,000 and 300,000 µg BPA/kg /day dose groups. In contrast, [21] observed a significant positive association between increased urine BPA concentration and higher prolactin and progesterone levels in a study done from BPA exposed and unexposed factories in China on 106 exposed and 250 unexposed female workers. In addition, a positive association between urine BPA and estrogen was observed among exposed workers with borderline significance, while a statistically significant inverse association between urine BPA and FSH was observed among unexposed group. These authors reported that BPA exposure was found to be linked to higher prolactin level among adult females. However, this finding is consistent with the results of the present study and other reported findings from *in vitro* and *in vivo* studies. *In vitro* studies showed that except for estrogen receptors alpha, beta, gamma, BPA can also bind to membrane estrogen receptor (mER), and these membrane bound receptors are capable of non-genomic steroid actions [22]. GH3/B6 pituitary cells, which express mER, respond to low level BPA exposure by producing a calcium flux which leads to PRL

release [23]. BPA can also induce prolactin gene expression and cell proliferation in both primary anterior pituitaries cells and GH3 cells [24]. In an animal study, injecting approximately 15 mg/(kg/day) of BPA into neonatal Fisher 344 rat pups resulted in an increase in serum prolactin levels [25]. Similarly, treatment of ovary-ectomized Wistar rats with BPA doses of 11–250 mg/kg per day induced hyperprolactinemia [26]. The same result was found when perinatal administration of BPA (0.05 mg/kg/day, 20 mg/kg/day) was used, the exposed animals of F1 females reached adulthood and became pregnant, and it induced alterations in serum progesterone and estrogen 68hormonal levels[27] The exposure to 500 µg/kg/day BPA in rats leads to an ovulation and infertility[28] Moreover, BPA may target GnRH neurons and as a result cause the decrease in GnRH mRNA expression[29] . The uterus responds to the changing hormone levels produced by the brain as well as the ovaries. This process is initiated in the hypothalamus through the production and release of GnRH, which leads to FSH and LH release from the anterior pituitary gland. As a result, oocyte development takes place in the ovaries, and estradiol is produced from the growing ovarian follicle. BPA can affect the hypothalamic system. The hypothalamic-pituitary ovarian axis controls the ability of the mammalian female to ovulate and to prepare the reproductive organs to support potential pregnancy. BPA exposure resulted in the decrease of the reproductive capacity and delay or elimination of puberty [31]. BPA affects ovarian steroidogenesis by modulating the expression of key steroidogenic enzymes. For example, BPA decreases aromatase (CYP19A1) expression and E2 production in human granulosa cells [32]. In mice, BPA inhibits P4, testosterone (T) and E2 synthesis by decreasing the expression of steroidogenic acute regulatory protein (Star), 3β-hydroxysteroid dehydrogenase (Hsd3b1) and 17α- hydroxylase (Cyp17a1) [33] . In rats, however, BPA increases P and T synthesis, as well as the expression of Star, cholesterol side-chain cleavage enzyme (Cyp11a1) and Cyp17a1, but decreases E2 synthesis and Cyp19a1 expression (Zhou *et al.*, 2008). In pigs, BPA increases basal and FSH-induced P4 synthesis, whereas it decreases FSH-induced E2 synthesis [34] Histopathological changes in rat ovaries are consistent with Adult female rats exposed to BPA had atretic follicles, which were characterized by the initial elimination of granulosa cells proximal to the antrum, pyknotic nuclei, and remnants of mitochondrial and plasma membranes [35]. The presence of atretic follicles may be attributed to decreased estradiol concentration as reported by [36]. The present study has shown that orally administered BPA at 25 µg/kg/day caused significant decrease in the reproductive hormones, FSH, LH, Testosterone, with an elevated level of prolactin. The toxic effect of BPA on the male reproductive functions is 69 well defined in animal models and demonstrated by physiological changes throughout foetal, pubertal and adult life of male rats [37-39]. An *in vivo* study showed that when low doses of BPA were given to rats via oral administration an impairment of spermatogenesis caused by the reduction of reproductive hormones serum level (FSH, LH, GnRH) and stopping germ cells meiosis process, thus activating the apoptosis pathway in germ cells. BPA administration reduces testosterone biosynthesis and secretion, thus inhibiting the activity of GnRH neurons, and lowering the expression of steroidogenic enzymes. Consequently, a decline of testosterone levels and a reduction in spermatozoa concentration was seen [40]. BPA effect on testosterone level has been tested on prepubertal rats after subcutaneously administered doses of 0, 20, 100, and 200 mg/kg/day, for six weeks. This study has demonstrated the decrease in testosterone levels only after higher doses (100 and 200 mg/kg/day) [41]. Other studies have shown that BPA inhibits steroidogenesis in the rat testis and reduces testosterone secretion, thus inhibiting the activity of GnRH neurons, and lowering the

expression of steroidogenic enzymes [42-43] . Spermatogenesis is dependent on a well-orchestrated hormonal environment. Leydig cells stimulated by LH provide the local production of testosterone, and Sertoli cells stimulated by FSH provide the local production of estradiol. In addition, Sertoli cells maintain the spermatogonial stem cells responsible for the continuity of spermatogenesis [44] . In the present study, BPA exposure caused an imbalance in these hormones, which may have contributed to defects in spermatogenesis and sperm maturation. Using prepubertal rats as an experimental model,[45] also reported a dose-dependent reduction in testosterone and LH serum concentrations (20, 100 and 200mg BPA/kg/day). The reduction in the LH serum concentration may be directly responsible for the reduction in testosterone production by Leydig cells in BPA-treated animals. In the seminiferous tubules, testosterone is carried by androgen binding protein (ABP) through the testis toward the epididymis. Testosterone is converted to dihydrotestosterone (DHT) by 5-alpha reductase enzyme [46-47]. The androgenic 70activity of DHT is two-fold higher than testosterone, and the epididymis is highly dependent on androgens to complete its transport and storage of spermatozoa prior to ejaculation [46-47]. Therefore, the reduction in testosterone observed in the BPA-treated animals can affect these processes and may be at least partly responsible for the alterations observed in the spermatozoa of these rats.

5. Conclusion:

In a comprehensive study conducted on both male and female rats, a distinct decrease in reproductive hormone levels was prominently noted when exposed to Bisphenol A (BPA) at a dosage of 25 µg/kg per day. Interestingly, despite this suppression in hormonal activity, one notable exception was found in the case of Prolactin, which exhibited an unusual increase compared to the baseline levels. Furthermore, the repercussions of BPA exposure seemed to manifest differently in males and females. Male rats displayed concerning signs of seminiferous epithelium degeneration, which led to a significant decline in sperm production. Conversely, female rats exhibited notable abnormalities in their ovarian structure, characterized by the absence of healthy antral follicles, the enlargement of cystic follicles, and the presence of atretic follicles. These findings shed light on the detrimental effects of BPA on the reproductive systems of both male and female rats, emphasizing the urgent need for further investigations into the impact of this chemical on overall reproductive health.

References

1. Gregoraszczyk EL, Ptak A (2013). Endocrine-disrupting chemicals: some actions of POPs on female reproduction. *Int J Endocrinol.* 2013; 2013:828532.
2. Cao XL, Zhang J, Goodyer CG, Hayward S, Cooke GM, Curran IH (2012). Bisphenol A in human placental and fetal liver tissues collected from greater Montreal area (Quebec) during 1998-2008. *Chemosphere.* 2012;89(5):505–11.
3. Lee J, Choi K, Park J, Moon HB, Choi G, Lee JJ, et al (2018). Bisphenol A distribution in serum, urine, placenta, breast milk, and umbilical cord serum in a birth panel of mother-neonate pairs. *Sci Total Environ.* 2018; 626:1494–501.
4. Wang Y, Zhu Q, Dang X, He Y, Li X, Sun Y (2017). Local effect of Bisphenol A on the estradiol synthesis of ovarian granulosa cells from PCOS. *Gynecol Endocrinol.* 2017;33(1):21–5.
5. Santos-Silva AP, Andrade MN, Pereira-Rodrigues P, Paiva-Melo FD, Soares P, Graceli JB, et al (2017). *Frontiers in*

- endocrine disruption: impacts of organotin on the hypothalamus-pituitary-thyroid axis. *Mol Cell Endocrinol.* 2017.
6. Cocci P, Capriotti M, Mosconi G, Palermo FA (2017). Effects of endocrine disrupting chemicals on estrogen receptor alpha and heat shock protein 60 gene expression in primary cultures of loggerhead sea turtle (*Caretta caretta*) erythrocytes. *Environ Res.* 2017; 158:616–24.
 7. Engel A, Buhre T, Imber F, Jessel S, Seidel A, Volkel W, et al (2017). Agonistic and antagonistic effects of phthalates and their urinary metabolites on the steroid hormone receptors ERalpha, ERbeta, and AR. *Toxicol Lett.* 2017;277: 54–63.
 8. Rattan S, Zhou C, Chiang C, Mahalingam S, Brehm E, Flaws JA (2017). Exposure to endocrine disruptors during adulthood: consequences for female fertility. *J Endocrinol.* 2017;233(3): R109–29.
 9. Aviva Systems Biology (2010-2019), Address: 7700 Ronson Rd Jury Classroom / Gallery, San Diego, CA 92111, United States.
 10. Solanki M, Visscher D (2020). Pathology of breast cancer in the last half century. *Human Pathology.* 2020 Jan 1;95: 137-48.
 11. Pandiyan P, Vijayakumar M, Chandrabose GS (2023). Statistical Methods (A Test-Book Written Completely On Modern Lines For Undergraduate & Postgraduate Courses In Statistics, Agriculture, Related Science Courses and Medical.). *BFC Publications*; 2023 May 5.
 12. Srivastava S, Dhagga N (2019). Dose exposure of Bisphenol-A on female Wistar rats fertility. *Hormone Molecular Biology and Clinical Investigation.* 2019 May 27;38(2):20180061.
 13. Hamdy H, Yahia D, Afifi S, Salem DA (2018). Endocrine disruption induced by bisphenol A in young and adult female Sprague Dawley rats. *Comparative Clinical Pathology.* 2018 Jul;27: 967-74.
 14. Mahalingam S, Ther L, Gao L, Wang W, Ziv-Gal A, Flaws JA (2017). The effects of in utero bisphenol A exposure on ovarian follicle numbers and steroidogenesis in the F1 and F2 generations of mice. *Reproductive toxicology.* 2017 Dec 1;74: 150-7.
 15. Amar S, Binet A, Tétéau O, Desmarchais A, Papillier P, et.al (2020). Bisphenol S impaired human granulosa cell steroidogenesis in vitro. *International Journal of Molecular Sciences.* 2020 Mar 6;21(5):1821.
 16. Stocco C (2008). Aromatase expression in the ovary: hormonal and molecular regulation. *Steroids.* 2008 May 1;73(5):473-87.
 17. Patel S, Brehm E, Gao L, Rattan S, Ziv-Gal A, Flaws JA (2017). Bisphenol A exposure, ovarian follicle numbers, and female sex steroid hormone levels: results from a CLARITY-BPA study. *Endocrinology.* 2017 Jun 1;158(6):1727-38.
 18. Stocco DM (2001). StAR protein and the regulation of steroid hormone biosynthesis. *Annual review of physiology.* 2001 Mar;63(1):193-213.
 19. Goyeneche AA, Harmon JM, Telleria CM (2006). Cell death induced by serum deprivation in luteal cells involves the intrinsic pathway of apoptosis. *Reproduction.* 2006 Jan 1;131(1):103-11.
 20. Camacho L, Lewis SM, Vanlandingham MM, Olson GR, Davis KJ,(2019). A two-year toxicology study of bisphenol A (BPA) in Sprague-Dawley rats: CLARITY-BPA core study results. *Food and Chemical Toxicology.* 2019 Oct 1;132: 110728.
 21. Liang H, Xu W, Chen J, Shi H, Zhu J,(2017). The association between exposure to environmental bisphenol A and gonadotropic hormone levels among men. *PloS one.* 2017 Jan 13;12(1): e0169217.
 22. Caserta D, Di Segni N, Mallozzi M, Giovanale V, Mantovani A, Marci R, Moscarini M (2014). Bisphenol A and the female reproductive tract: an overview of recent laboratory evidence and epidemiological studies. *Reproductive Biology and Endocrinology.* 2014 Dec; 12:1-0.
 23. Jeng YJ, Watson CS (2011). Combinations of physiologic estrogens with xenoestrogens alter ERK phosphorylation profiles in rat pituitary cells. *Environmental health perspectives.* 2011 Jan;119(1):104-12.
 24. Rebuli ME, Cao J, Sluzas E, Delclos KB, Camacho L, et.al (2014). Investigation of the effects of subchronic low dose oral exposure to bisphenol A (BPA) and ethinyl estradiol (EE) on estrogen receptor expression in the juvenile and adult female rat hypothalamus. *Toxicological Sciences.* 2014 Jul 1;140(1):190-203.
 25. Hernandez ME, Soto-Cid A, Rojas F, Pascual LI, Aranda-Abreu GE, et.al (2006). Prostate response to prolactin in sexually active male rats. *Reproductive Biology and Endocrinology.* 2006 Dec; 4:1-2.
 26. Ahsan N, Ullah H, Ullah W, Jahan S (2018). Comparative effects of Bisphenol S and Bisphenol A on the development of female reproductive system in rats; a neonatal exposure study. *Chemosphere.* 2018 Apr 1;197 :336-43.
 27. Martínez-Peña AA, Rivera-Baños J, Méndez-Carrillo LL, Ramírez-Solano MI, Galindo-Bustamante A, et.al (2017). Perinatal administration of bisphenol A alters the expression of tight junction proteins in the uterus and reduces the implantation rate. *Reproductive toxicology.* 2017 Apr 1;69: 106-20.
 28. Mahalingam S, Ther L, Gao L, Wang W, Ziv-Gal A, Flaws JA (2017). The effects of in utero bisphenol A exposure on ovarian follicle numbers and steroidogenesis in the F1 and F2 generations of mice. *Reproductive toxicology.* 2017 Dec 1;74: 150-7.
 29. Rebuli ME, Cao J, Sluzas E, Delclos KB, Camacho L, et.al (2014). Investigation of the effects of subchronic low dose oral exposure to bisphenol A (BPA) and ethinyl estradiol (EE) on estrogen receptor expression in the juvenile and adult female rat hypothalamus. *Toxicological Sciences.* 2014 Jul 1;140(1):190-203.
 30. Liu JC, Yan ZH, Li B, Yan HC, De Felici M, Shen W (2021). Di (2-ethylhexyl) phthalate impairs primordial follicle assembly by increasing PDE3A expression in oocytes. *Environmental Pollution.* 2021 Feb 1;270: 116088.
 31. Presunto M, Mariana M, Lorigo M, Cairrao E (2023). The effects of bisphenol A on human male infertility: A review of current epidemiological studies. *International journal of molecular sciences.* 2023 Aug 4;24(15):12417.
 32. Shi J, Liu C, Chen M, Yan J, Wang C, Zuo Z, He C (2021). The interference effects of bisphenol A on the synthesis of steroid hormones in human ovarian granulosa cells. *Environmental Toxicology.* 2021 Apr;36(4):665-74.
 33. Mahalingam S, Ther L, Gao L, Wang W, Ziv-Gal A, Flaws JA (2017). The effects of in utero bisphenol A exposure on ovarian follicle numbers and steroidogenesis in the F1 and F2

- generations of mice. *Reproductive toxicology*. 2017 Dec 1;74:150-7.
34. Mansur A, Adir M, Yerushalmi G, Hourvitz A, Gitman H, et.al (2016). Does BPA alter steroid hormone synthesis in human granulosa cells in vitro?. *Human Reproduction*. 2016 Jul 1;31(7):1562-9.
 35. Rodgers RJ, Irving-Rodgers HF (2010). Morphological classification of bovine ovarian follicles. *Reproduction*. 2010 Feb 1;139(2):309.
 36. Mahalingam S, Ther L, Gao L, Wang W, Ziv-Gal A, Flaws JA (2017). The effects of in utero bisphenol A exposure on ovarian follicle numbers and steroidogenesis in the F1 and F2 generations of mice. *Reproductive toxicology*. 2017 Dec 1;74:150-7.
 37. Vom Saal FS, Vandenberg LN (2021). Update on the health effects of bisphenol A: overwhelming evidence of harm. *Endocrinology*. 2021 Mar 1;162(3): bqaa171.
 38. Ahbab MA, Barlas N, Karabulut G (2017). The toxicological effects of bisphenol A and octylphenol on the reproductive system of prepubertal male rats. *Toxicology and industrial health*. 2017 Feb;33(2):133-46.
 39. Ullah H, Ambreen A, Ahsan N, Jahan S (2017). Bisphenol S induces oxidative stress and DNA damage in rat spermatozoa in vitro and disrupts daily sperm production in vivo. *Toxicological & Environmental Chemistry*. 2017 Jul 3;99(5-6):953-65.
 40. Liu X, Wang Z, Liu F (2021). Chronic exposure of BPA impairs male germ cell proliferation and induces lower sperm quality in male mice. *Chemosphere*. 2021 Jan 1;262: 127880.
 41. Lv Y, Li L, Fang Y, Chen P, Wu S, et.al (2019). In utero exposure to bisphenol A disrupts fetal testis development in rats. *Environmental Pollution*. 2019 Mar 1;246: 217-24.
 42. La Merrill MA, Vandenberg LN, Smith MT, Goodson W, Browne P,(2020). Consensus on the key characteristics of endocrine-disrupting chemicals as a basis for hazard identification. *Nature Reviews Endocrinology*. 2020 Jan;16(1):45-57.
 43. Cariati F, D'Uonno N, Borrillo F, Iervolino S, Galdiero G, Tomaiuolo R (2019). Bisphenol a: an emerging threat to male fertility. *Reproductive Biology and Endocrinology*. 2019 Dec; 17:1-8.
 44. O'Shaughnessy PJ (2014). Hormonal control of germ cell development and spermatogenesis. In *Seminars in cell & developmental biology* 2014 May 1 (Vol. 29, pp. 55-65). *Academic Press*.
 45. Maamar MB, Lesné L, Desdoits-Lethimonier C, Coiffec I, Lassarguère J,et.al (2015). An investigation of the endocrine-disruptive effects of bisphenol a in human and rat fetal testes. *PLoS One*. 2015 Feb 23;10(2):e0117226.
 46. Robaire B, Hamzeh M (2011). Androgen action in the epididymis. *Journal of andrology*. 2011 Nov 12;32(6):592-9.
 47. Domeniconi RF, Souza AC, Xu B, Washington AM, Hinton BT (2016). Is the epididymis a series of organs placed side by side?. *Biology of reproduction*. 2016 Jul 1;95(1):10-.

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